

Maryland Department of Health & Mental Hygiene

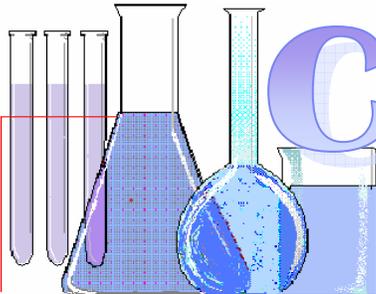
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Anthony G. Brown
Lt. Governor

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CRITICAL LINK



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Narrowing the Diagnostic Window: Identifying Acute HIV Infections

Acute Infections and HIV transmission

The initial phase of HIV disease is referred to as the acute phase. After HIV enters the body it replicates to high levels until host immune defenses are activated and reduce the initially high viral burdens (viral loads) to lower chronic baseline levels that persist for years. The duration of the acute phase is estimated to range from 54 to 62 days^{1,2} with viral loads spiking during the first two to five weeks after acquisition of the infection.^{1,3,4,5} The acute phase is not only characterized by high HIV viral loads in the plasma (blood) but also by viral shedding from the genital tract. Viral shedding in the genital tract has been associated with higher rates of transmission per exposure than later phases of infection.^{1,3} It has been estimated that nearly half of all HIV transmissions can be attributed to acutely infected individuals who are highly infectious and unaware of their disease status.^{3,6,7}

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Gonococcus (GC) Media Report

Over the past 20 months, the Laboratories Administration's Public Health Microbiology Division conducted investigations to determine why there has been a significant increase in the number of *Neisseria gonorrhoeae* cultures that are being reported out as "unsatisfactory" (unsat) due to overgrowth by normal flora. During this period, the local health departments (LHDs) have been involved and worked with the State Laboratory in studying this problem.

Beginning in April 2005, the Public Health Microbiology Division reviewed and studied a number of possible causes and solutions. These include comparing media from different manufacturers, requesting and comparing reformulated media, and looking into such variables as specimen collection errors, transport temperature, delivery times, individual plating and incubating procedures, and incubation times. While these variables accounted for specific problems at certain times in particular clinics, all were ruled out as the cause of the overgrowth problem that was occurring across the State.

Various LHD clinics were asked in the past year to use a particular type of culture system (i.e., Transgrow™ bottles, Jembec™ plates, and modified Thayer-Martin agar [MTM II] plates). Between January 2005 and July 2007 Transgrow™ and Jembec™ plates were found in any given month to have overgrowth rates as high as 18% and 7%, respectively. However, using these media did not resolve the overgrowth problem. After 20 months of investigating this problem, and finding no smoking gun, it appears that the normal flora from the throat, vagina, rectum, and urethra of many clinic patients may simply be expressing a naturally increased resistance to the antibiotics in GC culture media.

Nevertheless, the studies have proved useful. They have allowed the Public Health Microbiology Division to compile a list of important tips for LHD clinics collecting and plating GC speci-

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(Continued from page 1)

*Narrowing the Diagnostic Window:
Identifying Acute HIV Infections*

Identifying Acute HIV Infections

For more than two decades HIV diagnostic testing primarily has relied on the detection of HIV specific antibodies that are produced as part of the host immune response to the infection. However, using existing HIV screening tests, detectable levels of HIV specific antibodies cannot be demonstrated in blood for at least one to two weeks after HIV genetic material (RNA) can be detected in the blood by nucleic acid amplification testing (NAAT).^{2,10} Therefore HIV-1 NAAT has become the method of choice to identify acute HIV-1 infections. The early stage of an acute HIV infection is known as the "window phase." The window phase is characterized by high levels of HIV in the blood (viremia) that can be detected by HIV-1 NAAT prior to the production of HIV specific antibodies (seroconversion) by the acutely infected individual. The duration of the window phase is determined by the relative sensitivity of HIV antibody screening (Enzyme Immuno-Assay: EIA or rapid point-of-care tests). Newer, third-generation EIA's based on recombinant or synthetic peptide antigens generally can detect HIV specific antibodies days or weeks earlier than EIA's based on whole viral lysate antigens that were originally developed in the 1980's or early 1990's.⁹ Additionally, newer, third-generation EIA's are more sensitive than the traditional HIV-1 western blot confirmatory assay and most of the rapid point-of-care screening tests. Many third-generation EIA reactive specimens from acutely HIV-1 infected individuals can be initially non-reactive in rapid screening tests or, if reactive in newer generation screening immunoassays, cannot be confirmed as HIV-1 antibody-positive by less sensitive traditional western blot (WB) testing. This effectively prolongs the time of the window phase until HIV-1 seroconversion can be confirmed.

HIV-1 NAAT of Blood Donors

Recognizing the importance of identifying acute HIV infections to prevent transmission via HIV infected blood products, the blood banking centers in the United States began using HIV-1 NAAT to screen pools of 16-24 HIV antibody negative specimens in March 1999.¹¹ If a pool is found to contain HIV RNA, it is deconstructed and each specimen making up the pool is tested individually to identify the specimen(s) from the acutely infected donor(s). Pooling protocols allow fewer tests to be performed per specimen and thus reduce the high cost of performing HIV-1 NAAT when screening large numbers of specimens. Due to the low prevalence of HIV infections in blood donors, the identification of acute HIV infections by HIV-1 NAAT rarely occurs in this population. On average, one acutely HIV-1 infected donor is identified by HIV-1 NAAT for every 3.1 million donated units tested.¹¹

HIV-1 NAAT in Public Health Settings

Starting in North Carolina in 2002, state and local public health laboratories began to utilize NAAT to identify acute HIV-1 infections within their testing populations. Testing programs in public health settings designed to detect acute infections by HIV-1 NAAT currently exist in San Francisco, King County in Washington State, Los Angeles, Maryland, and other sites.⁸ During the

first year of HIV-1 NAAT in North Carolina, 23 acute HIV-1 infections were identified by testing 109,250 persons.¹² The increased rate of detecting acute HIV-1 infections in public health testing populations compared to blood donors, reflects larger numbers of individuals within public health testing populations engaging in high risk behaviors (i.e., unprotected sex with multiple partners, and IV drug abuse) and the use of less sensitive, first generation viral lysate-based screening EIA's. HIV-1 NAAT screening in support of public health HIV prevention programs has demonstrated potential utility in several ways by: 1) Increasing the diagnostic yield of testing programs above traditional HIV antibody testing; 2) Identifying and describing nascent outbreaks of HIV transmission; and 3) Providing opportunities to rapidly intervene to reduce potential HIV transmissions from acutely infected individuals. The initial study in North Carolina projected that HIV-1 NAAT protected 48 sexual partners from high-risk exposures to individuals who were identified as acutely HIV-1 infected.¹²

HIV-1 NAAT at the MD DHMH Laboratory

Since October 2004, the DHMH Retrovirology Laboratory has utilized HIV-1 NAAT to identify individuals who are acutely infected with HIV-1 as part of a comprehensive HIV testing algorithm. The Retrovirology Laboratory validated cost-effective, real-time HIV-1 PCR assays that target the LTR and gag genes of HIV-1. These assays demonstrated a reproducible lower limit of detection at approximately 2,000 HIV-1 RNA copies/ml per single member of a 20-specimen, serum/plasma pool. The DHMH Retrovirology Laboratory uses HIV-1 NAAT in two different testing strategies to quickly identify potentially acute HIV-1 infections in our testing populations. One testing strategy involves testing pools of 20 serum/plasma specimens that have been screened negative for HIV antibodies by two screening EIA's. The other strategy is to test individual diagnostic specimens that were reactive in screening HIV EIA's but cannot be confirmed as HIV-1 antibody-positive by HIV-1 WB testing. When potential acute HIV-1 infections are detected by NAAT, the health care provider is immediately contacted by the laboratory and urged to quickly obtain follow-up specimens for additional testing to resolve the patient's HIV infection status.

Results of Testing HIV Serology Specimens with Inconclusive Results

From October 2004 through August 2007, 133,601 serum/plasma specimens (excluding cadaveric specimens) were submitted to the DHMH Retrovirology Laboratory for routine HIV antibody testing. A total of 398 HIV EIA reactive specimens that could not be confirmed as HIV-1 antibody positive by WB testing were individually tested by real-time PCR (NAAT). Thirty-four (34) specimens (8.54% of total tested) were found to contain HIV-1 RNA by HIV-1 NAAT. The 34 RNA positive specimens were identified from 29 individuals (27 suspected HIV-1 seroconversion and two end-stage AIDS patients). HIV-1 viral loads were determined by branched DNA (bDNA) from 12 of 27 suspected HIV-1 seroconversion patients that had specimens with sufficient volumes to permit testing. The HIV-1 viral loads in these specimens ranged from 287 to >500,000 copies/ml (mean: 197,658 copies/ml). Nineteen of the 27 suspected seroconversion cases were subsequently confirmed as HIV-1 antibody positive by serological testing of follow-up specimens. The remaining eight individuals remain lost to follow-up. Five of 19 confirmed HIV-1 seroconversion cases initially were HIV-1/HIV-2

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Narrowing the Diagnostic Window: Identifying Acute HIV Infections

EIA reactive and HIV-1WB negative. In recognition of the increased sensitivity of newer generations of HIV screening EIA's relative to conventional HIV-1 WB confirmatory testing, our data demonstrated the utility of individual HIV-1 NAAT to identify acute HIV-1 infections in patients with inconclusive serology results. The high HIV-1 viral loads present during most of the acute phase of HIV infections allow available diagnostic sera to be quickly used for HIV-1 NAAT in lieu of ideal plasma specimens that would have to be collected later. The logistical difficulties that can be encountered when collecting and testing a follow-up plasma specimen could delay by days or weeks the recognition of acute HIV infections.

Results of Testing HIV EIA Negative Specimens

Since October 2004 over 125,000 HIV EIA antibody-negative specimens have been tested by HIV-1 NAAT in pools of 20 specimens by the DHMH Retrovirology Laboratory. To date only two EIA HIV-1 NAAT positive specimens have been identified. One of these patients was later confirmed as HIV-1 infected by follow-up serological testing, and the other remains lost to follow-up. The lower yield of acute HIV-1 infections using this testing strategy can be attributed to several factors. The DHMH laboratory uses a newer third generation HIV-1/HIV-2 screening EIA that is more sensitive in detecting early antibody responses that are mounted during early stages of acute HIV infections in comparison to older viral-lysate-based screening EIAs that have been used by other public health laboratories when performing HIV-1 NAAT testing. Five additional HIV-1 NAAT positive confirmed cases and two HIV-1 NAAT positive suspected cases of HIV-1 seroconversion were identified by the DHMH during this time period. They were exclusively reactive in the third generation (synthetic peptide based) EIA but were antibody negative when tested in parallel in the less sensitive viral lysate EIA. Secondly, the DHMH Laboratory tests all specimens submitted for HIV antibody testing and does not selectively target clinics with more at-risk clients where the yield of acutely infected individuals is higher. Finally, many high-risk populations are now tested for HIV antibodies using point-of-care rapid tests, and blood specimens from these individuals that could be used to perform HIV-1 NAAT are not routinely submitted to the DHMH Laboratory to identify possible HIV antibody-negative acute infections.

Concluding Remarks

In conclusion, HIV-1 NAAT has improved the ability of laboratories to quickly identify potentially acute HIV infections effectively narrowing the diagnostic "window phase" before HIV-1 antibodies can be conclusively identified by conventional serological testing. HIV-1 NAAT is strongly recommended when testing at-risk individuals in testing populations with high HIV prevalence where the incidences of acute HIV infections are projected to occur more frequently. The early identification of acute HIV-1 infections not only benefits the individual patient by quickly implementing anti-viral therapies but also can target HIV prevention strategies through behavior modification programs and immediate partner identification and notification.

This article written by Dr. Robert A. Myers



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Department of Health & Mental Hygiene

201 W. Preston Street
Baltimore, MD 21201
(Phone 410-767-6909)

Critical Link: Production Manager
Georgia Corso

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Questions concerning technical content of this
newsletter may be referred to
Dr. Jack DeBoy at 410-767-6100

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- ¹ *J. Infect. Dis.* 2004;189(10):1785-1792.
- ² *AIDS.* 2003;17: 1871-1879.
- ³ *J. Infect. Dis.* 2005; 191:1403-1409.
- ⁴ *J. Acquir. Immune Defic. Syndr. Hum. Retrovirol.* 14:249-258. 1997
- ⁵ *AIDS.* 2005;19:1372-1319.
- ⁶ *Am. J. Public Health.* 1997;87:1928-1930.
- ⁷ *JAIDS.* 2005;38(5):531-537.
- ⁸ ASTHO Issue Brief: Acute HIV Infections An Opportunity to Enhance Primary Prevention 2006:1-18 available at www.astho.org/pubs/AcuteHIVInf.pdf.
- ⁹ *J. Clin. Micro.* 2006;44(5):1856-1858.
- ¹⁰ *JAMA* 2003;289:959-962.
- ¹¹ *N. Engl. J. Med.* 2004;351(8):760-768.
- ¹² *N. Engl. J. Med.* 2005;352(18):1873-1883.

mens (see Table 1). Also, based on input from the LHDs (e.g., preference and ease of use), several technical considerations (e.g., area of plate), and use by public health laboratories in other states, the Laboratories Administration has decided to provide all LHDs with the MTM II culture system (media plate, Dacron™ swab, CO₂-generating tablet, and resealable polyethylene bag) as soon as procurement and delivery arrangements can be worked out with the manufacturer.

The scientists in the Public Health Microbiology Division will provide instructions (see Tables 1, 2, and 3) with initial shipments of MTM II media to all clinic sites. Laboratory staff will also be available to answer any questions from LHD staff by phone (410-767-6132). We will also continue a monthly review of the numbers of specimens that are reported unsatisfactory due to overgrowth.

This article written by Dr. Jack DeBoy and Shenia Young.

Processing Specimens to Culture for *Neisseria gonorrhoeae* Using MTM II Agar

Table 1. Procedure for using the Modified Thayer-Martin (MTM II) agar plate system when culturing specimens from the throat, vagina, rectum or urethra for *Neisseria gonorrhoeae*.^a

1. Process the specimen as soon as it is collected.
2. Roll swab directly on the medium in a large "Z" (to provide adequate exposure of the swab to the medium for transfer of organisms.)
3. Label plates legibly with patient's name (don't use China markers—their marking smudges and rubs off when wet.)
4. Place inoculated plates in the resealable polyethylene bag (one specimen per patient with accompanying lab slip.)
5. Cut off the corner of one foil-wrapped CO₂ tablet to expose the tablet and place it in the bag. DO NOT REMOVE THE TABLET FROM THE FOIL POUCH.
6. Expel excess air from the bag and completely seal the bag.
- 7a. If an incubator is available, incubate the plates in an inverted (medium facing down) position at 35°C until picked up by courier.
- 7b. If an incubator is not available, invert the plates and hold them at room temperature until picked up by courier. Do not refrigerate after inoculating.
8. When packing plates for transport, keep them inverted and place in a suitable container that will protect them from extreme heat or cold.

^aAdapted from: "Quality Control/Product Information". May, 1997. Modified Thayer-Martin (MTM II) Agar, Product Info. No. P21567 Rev. 05. BBL® Quality Control and Product Information Manual for Plated Media, Becton Dickinson Microbiology Systems, Cockeysville, MD, pp. 1-5 (0597 193 –0597 197).

Supplies:

1. MTM II plate
2. Dacron™ swab
3. Whirl-Pak resealable bag
4. CO₂ tablet in foil pouch

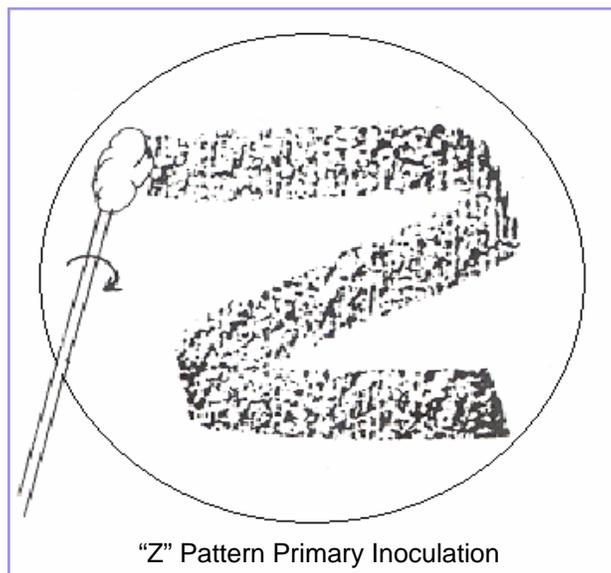


Table 2. Important tips (requirements) when plating and processing Modified Thayer-Martin (MTM II) agar plates to culture *Neisseria gonorrhoeae*.

1. Store plates under refrigeration upside down (media facing down.)
2. Discard any plate(s) with an expired expiration date or that exhibit growth prior to use (never use contaminated plates.)
3. Always allow plates to warm to room temperature before using (cold kills *N. gonorrhoeae*).
4. Use Dacron™-tipped swabs with plastic shafts (do not use cotton-tipped swabs, they may contain fatty acids that can interfere with the survival of some organisms. Also do not use calcium alginate-tipped swabs. They can be toxic for some strains of *N. gonorrhoeae*.)^a
5. Always allow surface of plates to dry before using (a wet surface hampers isolated colony formation.)
6. DO NOT CRUSH OR ADD WATER TO THE CO₂-GENERATING TABLET (CAUSES LOSS OF CO₂ AND POSSIBLE CONTAMINATION BY WATER.) MOISTURE FROM THE MEDIUM WILL ACTIVATE THE CO₂ TABLET.
7. Stack plates no more than four high in an incubator and at least one inch from the incubator wall and other stacks (uneven temperatures cause uneven growth.)
8. Don't incubate inoculated plates in the clinic longer than 24 hours (over-incubation leads to more growth of contaminating normal flora.)

^aManual of Clin. Micro. 2007 (9th ed.). Am. Soc. for Micro., p. 44.

Table 3. Tips for filling out laboratory request for gonococcus culture.

1. Fill in patient's name, DOB, case # if applicable, and last four digits of SSN.
2. Fill in date the specimen was collected.
3. Give specimen collection site (use specimen code.)
4. If incubated, indicate number of hours.
5. Make sure all lab slips contain submitter's information and patient information.

Laboratory Statistics

NS – Not Speciated
 NT – Non-Typeable
 VRE – Vancomycin Resistant
 SP – Species
 NG – No Growth

* This genus has recently been given a new genus name. The genus name in parenthesis is the old name.

** Formerly a part of the *Trichosporon beigeli* complex.

***Alpha streptococci other than *S. pneumoniae* and *Enterococcus*

REPORTED 10/01/07 - 10/31/07

ENTERIC BACTERIOLOGY

GENUS SEROVAR

SEX	AGE	#	JURISDICTION
CAMPYLOBACTER JEJUNI			
F	1	1	ALLEGANY
F	17	1	ANNE ARUNDEL
F	61	1	ANNE ARUNDEL
F	39	1	HARFORD
M	34	1	HARFORD
M	32	1	WORCESTER
M	59	1	BALTIMORE CITY
U	41	1	BALTIMORE CITY
M	32	1	OUT OF STATE
M	35	1	OUT OF STATE
M		1	UNKNOWN
CAMPYLOBACTER PRESENT			
M	41	1	HARFORD
M	32	1	WORCESTER
ESCHERICHIA COLI O157:H7			
M	65	1	BALTIMORE
F	80	1	CALVERT
M	10	1	TALBOT
M	57	1	WASHINGTON
M		1	BALTIMORE CITY
SALMONELLA AGONA			
U	1	1	BALTIMORE
M	1	1	CALVERT
M	40	1	OUT OF STATE
SALMONELLA BAREILLY			
U	5	1	PRINCE GEORGE'S
SALMONELLA BERTA			
F	40	1	ALLEGANY
SALMONELLA BOVISMORBIFICANS			
U	2	1	OUT OF STATE
SALMONELLA BRAENDERUP			
M	10	1	ANNE ARUNDEL
F	7	1	BALTIMORE
SALMONELLA DERBY			
U		1	OUT OF STATE
SALMONELLA ENTERITIDIS			
F	47	2	ANNE ARUNDEL
F	54	1	ANNE ARUNDEL
M		1	ANNE ARUNDEL
F	17	1	BALTIMORE
M		1	BALTIMORE
M	2	1	BALTIMORE
M	14	1	BALTIMORE
M	18	1	BALTIMORE
F	51	1	CALVERT
F	52	2	CALVERT

F	21	1	CHARLES
F	25	1	MONTGOMERY
F	59	1	MONTGOMERY
M	9	1	PRINCE GEORGE'S
F		1	BALTIMORE CITY
F	1	1	BALTIMORE CITY
F	2	1	BALTIMORE CITY
F	3	1	BALTIMORE CITY
F	10	1	BALTIMORE CITY
F	14	1	BALTIMORE CITY
F	20	1	BALTIMORE CITY
F	82	1	BALTIMORE CITY
M		2	BALTIMORE CITY
M	1	1	BALTIMORE CITY
M	6	1	BALTIMORE CITY
M	25	1	BALTIMORE CITY
M	41	1	BALTIMORE CITY
M	42	1	BALTIMORE CITY
U		1	BALTIMORE CITY
F	74	1	OUT OF STATE
M	9	1	OUT OF STATE
M	30	1	OUT OF STATE
M	39	1	OUT OF STATE
M	48	1	OUT OF STATE
SALMONELLA HEIDELBERG			
M	9	1	ST. MARY'S
SALMONELLA HVITTINGFOSS			
F	1	1	BALTIMORE
SALMONELLA I 13,23:b:-			
F	85	1	CARROLL
F	86	1	CARROLL
SALMONELLA I 6,7:k:-			
F	42	1	OUT OF STATE
SALMONELLA I 6,8:e,h:-			
F	32	1	FREDERICK
F	35	1	MONTGOMERY
SALMONELLA INFANTIS			
F	19	1	ANNE ARUNDEL
M	10	1	HARFORD
M		1	HOWARD
SALMONELLA ISRAEL			
F	24	1	MONTGOMERY
SALMONELLA IV 44:z4,z23:-			
M	5	1	ANNE ARUNDEL
U	46	1	OUT OF STATE
SALMONELLA JAVA			
U	6	1	ANNE ARUNDEL
F	3	1	BALTIMORE
M	63	1	PRINCE GEORGE'S
U	36	1	TALBOT
F	53	1	WASHINGTON
M	56	1	OUT OF STATE
SALMONELLA JAVIANA			
F	9	1	ANNE ARUNDEL
M		1	ANNE ARUNDEL
M	1	1	ANNE ARUNDEL
F	53	1	CAROLINE
F	3	2	WICOMICO
M	4	1	WICOMICO
F	48	1	WORCESTER
F	87	1	WORCESTER
F	35	1	BALTIMORE CITY
SALMONELLA LITCHFIELD			
U	5	1	ANNE ARUNDEL
SALMONELLA LOCKLEAZE			
U	62	1	OUT OF STATE
SALMONELLA MONTEVIDEO			
U	6	1	PRINCE GEORGE'S
F	16	1	OUT OF STATE

SALMONELLA MUENCHEN		
F	36	1
SALMONELLA NEWPORT		
F	1	1
U	28	1
F	8	1
M	49	1
M	17	1
F	46	1
M	1	1
M	18	1
M	61	1
F	24	1
U		1
F	2	1
U	1	1
U	4	1
SALMONELLA NORWICH		
M	1	2
SALMONELLA OHIO		
U	13	1
SALMONELLA ORANIENBURG		
M	35	1
F	8	1
M	1	1
U		1
SALMONELLA POMONA		
F		1
SALMONELLA SAINTPAUL		
U	8	1
F	24	1
U		1
SALMONELLA SANDIEGO		
M	9	1
M	50	1
U	23	1
SALMONELLA STANLEY		
F	44	1
SALMONELLA TENNESSEE		
F	70	1
SALMONELLA TYPHI		
F	10	1
M	33	1
U		1
SALMONELLA TYPHIMURIUM		
M	8	1
M	42	1
M	24	1
M	4	1
M	3	1
U	1	1
F	1	1
F	20	1
M	4	1
M	73	1
F		1
F	53	1
F		1
M	6	1
SALMONELLA TYPHIMURIUM VAR COPENHAGEN		
M		1
M		2
U	1	1
F	2	1
F	51	1
SALMONELLA UNTYPABLE		
F	38	1
F	64	1
M	33	1
M		1

CECIL
ANNE ARUNDEL
ANNE ARUNDEL
CAROLINE
CARROLL
DORCHESTER
FREDERICK
HOWARD
PRINCE GEORGE'S
TALBOT
WICOMICO
WICOMICO
WORCESTER
WORCESTER
OUT OF STATE
WICOMICO
MONTGOMERY
FREDERICK
MONTGOMERY
BALTIMORE CITY
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WASHINGTON
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OUT OF STATE
CARROLL
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HOWARD
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CARROLL
FREDERICK
MONTGOMERY
MONTGOMERY
PRINCE GEORGE'S
PRINCE GEORGE'S
ST. MARY'S
WASHINGTON
WASHINGTON
WASHINGTON
WICOMICO
WICOMICO
BALTIMORE CITY
BALTIMORE CITY
CAROLINE
MONTGOMERY
MONTGOMERY
TALBOT
BALTIMORE CITY
BALTIMORE
BALTIMORE
HARFORD
BALTIMORE CITY

M	65	1	BALTIMORE CITY
F	1	1	OUT OF STATE
U	1	1	OUT OF STATE
U	11	1	OUT OF STATE
SALMONELLA 4,5,12:i:-			
M	29	1	ALLEGANY
M		1	ANNE ARUNDEL
F	3	1	CARROLL
F	6	1	CHARLES
M	12	1	TALBOT
F		1	BALTIMORE CITY
M		1	BALTIMORE CITY
F	2	1	OUT OF STATE
SHIGELLA FLEXNERI II:3,4			
M	37	1	BALTIMORE CITY
SHIGELLA SONNEI			
M	52	1	ANNE ARUNDEL
F	2	1	PRINCE GEORGE'S
U	3	1	PRINCE GEORGE'S
F	2	1	OUT OF STATE
F	3	1	OUT OF STATE
M	68	1	OUT OF STATE
VIBRIO FLUVIALIS			
F	79	1	CALVERT
VIBRIO PARAHAEMOLYTICUS			
M	60	1	BALTIMORE
M	66	1	CHARLES
TOTAL		177	

ISOLATES – THROAT CULTURES

COUNTY	GROUP A ¹	NON-GROUP A
ALLEGANY	1	17
SOMERSET	1	9
WICOMICO	4	6
TOTAL	6	32

¹ *Streptococcus pyogenes*

BACTERIOLOGY IDENTIFICATIONS

Referrals

GENUS SPECIES	SOURCE	#	JURISDICTION
BACILLUS SPECIES			
	PLEURAL FLUID	1	CARROLL
HAEMOPHILUS INFLUENZAE NON-TYPEABLE			
	BLOOD	3	BALTIMORE CITY
	BLOOD	1	TALBOT
HAEMOPHILUS INFLUENZAE SEROTYPE F			
	BLOOD	1	BALTIMORE CITY
STAPHYLOCOCCUS AUREUS			
	GI TRACT	3	SOMERSET
	THIGH	2	WICOMICO
	WOUND	1	WICOMICO
TOTAL		12	

ISOLATES – MISCELLANEOUS

GENUS SPECIES	SOURCE	#	JURISDICTION
ACINETOBACTER LWOFFII			
	PUSTULE	1	SOMERSET
BACILLUS CEREUS			
	BLOOD	1	BALTIMORE CITY
BACILLUS SPECIES			
	ABSCESS	1	BALTIMORE
	BLOOD	1	BALTIMORE CITY

CLOSTRIDIUM DIFFICILE		
BLOOD	1	BALTIMORE CITY
CORYNEBACTERIUM SPECIES		
WOUND	1	BALTIMORE
BLOOD	1	BALTIMORE CITY
FOOT	1	CARROLL
LESION	1	CECIL
ENTEROBACTER CLOACAE		
BLOOD	2	BALTIMORE CITY
ENTEROCOCCUS FAECALIS		
BLOOD	1	BALTIMORE CITY
TOE	1	FREDERICK
ENTEROCOCCUS GALLINARUM		
BLOOD	1	BALTIMORE CITY
ESCHERICHIA COLI		
BLOOD	2	BALTIMORE CITY
GROIN	1	FREDERICK
TOE	1	FREDERICK
GENITAL	1	PRINCE GEORGE'S
EUBACTERIUM LENTUM		
BLOOD	1	BALTIMORE CITY
GARDNERELLA VAGINALIS		
CERVICAL	1	PRINCE GEORGE'S
VAGINAL	4	SOMERSET
KLEBSIELLA PNEUMONIEA		
BLOOD	2	BALTIMORE CITY
GROIN	1	FREDERICK
MICROCOCCUS SPECIES		
PUSTULE	1	SOMERSET
PROTEUS MIRABILIS		
EAR	1	BALTIMORE
SKIN	1	MONTGOMERY
PSEUDOMONAS AERUGINOSA		
EAR	1	BALTIMORE
BLOOD	2	BALTIMORE CITY
FOOT	1	FREDERICK
INCISION	1	FREDERICK
TOE	2	FREDERICK
PSEUDOMONAS LUTEOLA		
SKIN	1	MONTGOMERY
PSEUDOMONAS PUTIDA		
FOOT	1	FREDERICK
TOE	1	FREDERICK
SERRATIA MARCESCENS		
LIVER	1	BALTIMORE CITY
STAPHYLOCOCCUS AUREUS		
WOUND	1	ALLEGANY
BOIL	1	ANNE ARUNDEL
WOUND	1	ANNE ARUNDEL
ABSCESS	2	BALTIMORE
BOIL	2	BALTIMORE
MIDDLE EAR	1	BALTIMORE
WOUND	1	BALTIMORE
KNEE	1	BALTIMORE CITY
ABSCESS	3	BALTIMORE CITY
BLOOD	5	BALTIMORE CITY
CSF	1	BALTIMORE CITY
BREAST	1	CARROLL
FINGER	1	CARROLL
FOOT	1	CARROLL
LESION	1	CECIL
TOE	2	FREDERICK
FOOT	2	FREDERICK
SKIN	2	MONTGOMERY
WOUND	1	PRINCE GEORGE'S
STAPHYLOCOCCUS EPIDERMIDIS		
BLOOD	1	BALTIMORE CITY
STAPHYLOCOCCUS SPECIES		
BOIL	1	BALTIMORE
WOUND	1	BALTIMORE
BLOOD	2	BALTIMORE CITY
ABSCESS	1	CARROLL
ARM	1	CARROLL
TOE	7	FREDERICK

STENOTROPHOMONAS MALTOPHILIA		
AXILLARY	1	BALTIMORE
STREPTOCOCCUS ALPHA-HEMOLYTIC		
BLOOD	1	BALTIMORE CITY
STREPTOCOCCUS BETA-HEMOLYTIC GROUP A		
VAGINAL	5	ANNE ARUNDEL
BLOOD	1	BALTIMORE CITY
CSF	1	BALTIMORE CITY
STREPTOCOCCUS BETA-HEMOLYTIC GROUP B		
BLOOD	1	BALTIMORE CITY
VAGINAL	1	CECIL
VAGINAL	1	FREDERICK
VAGINAL	1	HOWARD
VAGINAL	4	MONTGOMERY
CERVICAL	7	PRINCE GEORGE'S
VAGINAL	15	PRINCE GEORGE'S
VAGINAL	6	SOMERSET
STREPTOCOCCUS NON-HEMOLYTIC		
FOOT	1	FREDERICK
STREPTOCOCCUS NON-HEMOLYTIC INTERMEDIUS		
BLOOD	1	BALTIMORE CITY
STREPTOCOCCUS NON-HEMOLYTIC SANGUIS		
BLOOD	1	BALTIMORE CITY
TOTAL	134	

SEXUALLY TRANSMITTED DISEASES

GENUS SPECIES	SEX	#	JURISDICTION
NEISSERIA GONORRHEAE			
	F	1	ALLEGANY
	M	1	ALLEGANY
	F	1	ANNE ARUNDEL
	M	3	ANNE ARUNDEL
	F	3	BALTIMORE
	M	3	BALTIMORE
	F	0	CAROLINE
	M	1	CAROLINE
	F	2	CARROLL
	M	1	CARROLL
	M	1	CECIL
	F	3	CHARLES
	M	1	CHARLES
	F	2	DORCHESTER
	M	1	DORCHESTER
	F	2	FREDERICK
	M	2	FREDERICK
	F	1	HARFORD
	F	2	HOWARD
	M	1	HOWARD
	F	1	KENT
	F	4	MONTGOMERY
	M	3	MONTGOMERY
	F	20	PRINCE GEORGE'S
	M	31	PRINCE GEORGE'S
	F	2	QUEEN ANNE'S
	M	0	QUEEN ANNE'S
	F	0	SOMERSET
	M	1	SOMERSET
	F	2	TALBOT
	M	0	TALBOT
	F	2	WASHINGTON CO
	M	1	WASHINGTON CO
	F	4	WICOMICO
	M	8	WICOMICO
	F	1	BALTIMORE CITY
	M	8	BALTIMORE CITY
	F	1	OUT OF STATE
	M	1	OUT OF STATE
TOTAL		122	

CHLAMYDIA TRACHOMATIS

F	10	ALLEGANY
M	6	ALLEGANY
F	25	ANNE ARUNDEL
M	15	ANNE ARUNDEL
F	19	BALTIMORE
M	20	BALTIMORE
F	3	CALVERT
M	2	CALVERT
F	2	CAROLINE
F	7	CARROLL
M	2	CARROLL
F	2	CECIL
M	5	CECIL
F	11	CHARLES
M	7	CHARLES
F	3	DORCHESTER
M	3	DORCHESTER
F	12	FREDERICK
M	7	FREDERICK
F	10	HARFORD
M	10	HARFORD
U	1	HARFORD
F	6	HOWARD
M	2	HOWARD
F	4	KENT
M	2	KENT
F	29	MONTGOMERY
M	30	MONTGOMERY
F	63	PRINCE GEORGE'S
M	42	PRINCE GEORGE'S
F	2	QUEEN ANNE'S
M	4	QUEEN ANNE'S
M	1	ST. MARY'S
F	11	SOMERSET
M	2	SOMERSET
F	1	TALBOT
F	6	WASHINGTON
M	2	WASHINGTON
F	28	WICOMICO
M	20	WICOMICO
U	1	WICOMICO
F	1	WORCESTER
M	2	WORCESTER
F	11	BALTIMORE CITY
M	36	BALTIMORE CITY
F	4	OUT OF STATE
M	2	OUT OF STATE
TOTAL	494	

SYPHILIS SEROLOGY

M	1	ALLEGANY
M	6	ANNE ARUNDEL
U	1	ANNE ARUNDEL
F	4	BALTIMORE
M	6	BALTIMORE
F	16	BALTIMORE CITY
M	19	BALTIMORE CITY
U	2	BALTIMORE CITY
F	1	CARROLL
M	1	DORCHESTER
F	1	FREDERICK
M	2	FREDERICK
F	2	HARFORD
M	1	HARFORD
F	1	HOWARD
F	5	MONTGOMERY
M	4	MONTGOMERY
U	1	MONTGOMERY
F	9	PRINCE GEORGES
M	21	PRINCE GEORGES
F	2	WASHINGTON
M	1	WASHINGTON
F	4	WICOMICO
M	3	WICOMICO
TOTAL	114	

PENICILLIN RESISTANT GONORRHEA STATISTICS

REPORTED QUARTERLY
NO REPORT THIS MONTH

MYCOBACTERIOLOGY

GENUS SPECIES	SEX	AGE	#	JURISDICTION
MYCOBACTERIUM TUBERCULOSIS				
M		27	1	ANNE ARUNDEL
M		37	1	CHARLES
F		28	1	HOWARD
F		81	1	MONTGOMERY
M		22	1	MONTGOMERY
M		77	1	PRINCE GEORGE'S
M		50	1	BALTIMORE CITY
MYCOBACTERIUM TUBERCULOSIS COMPLEX				
F		35	1	BALTIMORE
F		41	1	BALTIMORE
F		76	1	BALTIMORE
M		32	1	CARROLL
M		37	1	CHARLES
M		49	1	CHARLES
F		75	1	FREDERICK
F		28	1	HOWARD
M		29	1	HOWARD
F		66	1	MONTGOMERY
M		22	2	MONTGOMERY
M		26	1	MONTGOMERY
F		23	1	PRINCE GEORGE'S
M		25	1	PRINCE GEORGE'S
M		47	1	PRINCE GEORGE'S
M		64	1	PRINCE GEORGE'S
M		17	1	SOMERSET
F		42	1	WASHINGTON
M		50	1	BALTIMORE CITY
M		58	2	BALTIMORE CITY
F		22	1	OUT OF STATE
F		72	1	OUT OF STATE
MYCOBACTERIUM ABSCESSUS				
F		49	1	BALTIMORE CITY
MYCOBACTERIUM AFRICANUM				
M		42	1	PRINCE GEORGE'S
MYCOBACTERIUM AVIUM COMPLEX				
M		35	1	ALLEGANY
M		78	1	BALTIMORE
F		35	1	CALVERT
F		69	1	CARROLL
F		46	1	CECIL
M		42	1	DORCHESTER
F		60	1	FREDERICK
M		77	1	FREDERICK
M		78	1	FREDERICK
F		61	1	HOWARD
F		83	1	HOWARD
M		29	1	HOWARD
M		77	1	OUT OF STATE
MYCOBACTERIUM FORTUITUM				
F		23	1	PRINCE GEORGE'S
M		49	1	OUT OF STATE
MYCOBACTERIUM GORDONAE				
M		31	1	MONTGOMERY
F		23	1	PRINCE GEORGE'S
F		65	1	PRINCE GEORGE'S
M		28	1	PRINCE GEORGE'S
M		54	1	PRINCE GEORGE'S
M		48	1	QUEEN ANNE'S
M		60	1	OUT OF STATE
MYCOBACTERIUM MARINUM				
M		60	1	ANNE ARUNDEL
TOTAL			56	

MYCOLOGY

GENUS SPECIES	SEX	AGE	#	JURISDICTION
ALTERNARIA SP				
F		56	1	FREDERICK
M		84	1	BALTIMORE CITY
ASPERGILLUS FUMIGATUS				
F		79	1	TALBOT
BIPOLARIS SP				
F		48	1	CAROLINE
BLASTOMYCES DERMATITIDIS				
M		67	1	BALTIMORE CITY
CANDIDA ALBICANS				
F		28	1	ANNE ARUNDEL
F		17	1	MONTGOMERY
M		49	1	MONTGOMERY
F		20	1	PRINCE GEORGE'S
F		21	1	PRINCE GEORGE'S
F		60	1	PRINCE GEORGE'S
M		62	1	PRINCE GEORGE'S
F		18	1	SOMERSET
F		20	1	SOMERSET
CANDIDA GLABRATA				
F		75	1	PRINCE GEORGE'S
M		67	1	BALTIMORE CITY
CANDIDA GUILLIERMONDII				
F		55	1	BALTIMORE
CANDIDA PARAPSILOSIS				
F		21	1	SOMERSET
CHRYSOSPORIUM SP				
M		58	1	BALTIMORE CITY
F		43	1	OUT OF STATE
CRYPTOCOCCUS NEOFORMANS				
M		54	1	BALTIMORE CITY
CURVULARIA SP				
F		48	1	CAROLINE
ENGYDONTIUM ALBUM				
F		64	1	TALBOT
FUSARIUM SP				
F			1	WICOMICO
GEOTRICHUM SP				
M		71	1	MONTGOMERY
GLIOCLADIUM SP				
M		4	1	TALBOT
MUCOR SP				
M		69	1	ALLEGANY
NOCARDIA ASTEROIDES COMPLEX				
F		41	1	BALTIMORE CITY
M		58	1	BALTIMORE CITY
PENICILLIUM SP				
M		75	1	PRINCE GEORGE'S
PITHOMYCES SP				
M		45	1	BALTIMORE CITY
RHIZOPUS ORYZAE				
F		37	1	CALVERT
SCEDOSPORIUM AIOSPERMUM				
F		89	1	ANNE ARUNDEL
TRICHOPHYTON RUBRUM				
M		50	1	BALTIMORE CITY
M		62	1	BALTIMORE CITY
TRICHOPHYTON SP				
M			1	BALTIMORE CITY
TRICHOPHYTON TONSURANS				
F		2	1	CALVERT
F		3	1	CALVERT
M		5	1	BALTIMORE CITY
TRICHOSPORON ASAHII				
M		74	1	BALTIMORE CITY
TOTAL			40	

MYCOBACTERIUM SUSCEPTIBILITY RESULTS

DURING OCTOBER, 2007, SUSCEPTIBILITY RESULTS ON 29 ISOLATES OF *M. TUBERCULOSIS* COMPLEX * WERE IDENTIFIED.

TOTAL: 11 DRUG RESISTANT STRAINS FOUND

#	COUNTY	DRUG
1	ANNE ARUNDEL	@ to STREPTOMYCIN
1 ^B	BALTIMORE	@ to PYRAZINAMIDE
1	CHARLES	@ to ISONIAZID and STREPTOMYCIN
2 ^A	CHARLES	@ to STREPTOMYCIN
2 ^A	HOWARD	@ to RIFAMPIN and RIFABUTIN
2 ^A	HOWARD	@ to STREPTOMYCIN
2 ^A	MONTGOMERY	@ to STREPTOMYCIN

^A Two isolates from the same patient

^B Probable *M. bovis*

@ RESISTANT

**Mycobacterium tuberculosis* complex consists of:

M. tuberculosis
M. bovis
M. bovis, BCG
M. africanum
M. microti
M. canettii

PARASITOLOGY

GENUS SPECIES	#	JURISDICTION
PROTOZOA		
BLASTOCYSTIS HOMINIS	1	BALTIMORE
	1	FREDERICK
	3	MONTGOMERY
	1	PRINCE GEORGE'S
ENDOLIMAX NANA	1	FREDERICK
	7	MONTGOMERY
	5	PRINCE GEORGE'S
ENTAMOEBIA COLI	2	FREDERICK
	7	MONTGOMERY
	8	PRINCE GEORGE'S
ENTAMOEBIA HARTMANNI	1	PRINCE GEORGE'S
GIARDIA LAMBLIA	3	MONTGOMERY
	5	PRINCE GEORGE'S
	2	ST. MARY'S
IODAMOEBIA BUTSCHLII	1	MONTGOMERY
	1	PRINCE GEORGE'S
TOTAL	49	
NEMATODES		
ENTEROBIUS VERMICULARIS	2	CARROLL
	1	KENT
HOOKWORM	1	FREDERICK
	1	PRINCE GEORGE'S
	3	WASHINGTON
TOTAL	8	
TREMATODES		
CLONORCHIS SINENSIS	1	PRINCE GEORGE'S
TOTAL	1	

ARTHROPOD IDENTIFICATION

NONE

TICK IDENTIFICATION

NONE



This micrograph depicts an egg of the parasitic trematode, or flat-worm, *Clonorchis sinensis*. In Prince George's County, a patient tested positive for this parasite in October. These eggs range in size from 27-35µm x 11-20µm. The operculum, at the smaller end of the egg, is convex, and rests on a visible "shoulder". At the opposite, larger abopercular end, a small knob or hook-like protrusion is often observed. (Source: CDC Public Health Image Library)

WATER MICROBIOLOGY

	# TESTED	# NON-COMPLIANT
COMMUNITY	2	0
NON-COMMUNITY	452	116
TOTAL	454	116

FOOD SAFETY

FOOD AND SHELLFISH MICROBIOLOGY

	# OF SAMPLES	NOTABLE PATHOGENS
FOOD	7	0
		# STANDARDS EXCEEDED *
CRABMEAT	0	0
		# STANDARDS EXCEEDED **
SHELLFISH	0	0
SHELLFISH GROWING WATERS	404	
TOTAL	411	0

STANDARDS

* CRABMEAT-FRESH
ESCHERICHIA COLI = LESS THAN 36 MPN/100 GRAM
 STANDARD PLATE COUNT = LESS THAN 100,000 PER GRAM

** SHELLFISH
 FECAL COLIFORMS = LESS THAN 230 MPN/100 GRAM
 STANDARD PLATE COUNT = LESS THAN 500,000 PER GRAM

VIRUS ISOLATION

ISOLATE	SEX	AGE	#	JURISDICTION
ECHOVIRUS	M	2	1	PRINCE GEORGE'S
SUBTOTAL			1	

HERPES SIMPLEX I

F	20	1	ALLEGANY
F	21	1	ALLEGANY
M	22	1	ALLEGANY
F	18	1	BALTIMORE
F	19	1	BALTIMORE
F	20	1	BALTIMORE
F	39	1	CARROLL
M	46	1	CARROLL
F	18	1	CECIL
M	47	1	CECIL
F	24	1	CHARLES
F	30	1	CHARLES
F	17	1	DORCHESTER
F	20	1	FREDERICK
F	25	1	FREDERICK
F	25	1	HARFORD
F	19	1	HOWARD
F	17	1	KENT
F	20	3	PRINCE GEORGE'S
F	23	1	PRINCE GEORGE'S
M	20	1	PRINCE GEORGE'S
F	20	1	BALTIMORE CITY
F	28	1	BALTIMORE CITY
M	29	1	BALTIMORE CITY
U	15	1	BALTIMORE CITY

SUBTOTAL 27

HERPES SIMPLEX II

F	27	1	ANNE ARUNDEL
F	29	1	ANNE ARUNDEL
F	40	1	ANNE ARUNDEL
M	49	1	ANNE ARUNDEL
F	19	2	BALTIMORE
F	22	2	BALTIMORE
F	25	1	BALTIMORE
F	33	1	BALTIMORE
F	43	1	BALTIMORE
M	20	1	BALTIMORE
F	16	1	CALVERT
M	20	1	CARROLL
M	24	1	CARROLL
M	46	1	CARROLL
F	47	1	CECIL
F	24	1	CHARLES
M	24	1	CHARLES
M	31	1	CHARLES
F	22	1	FREDERICK
F	38	1	FREDERICK
M	20	1	FREDERICK
F	21	2	MONTGOMERY
F	45	1	MONTGOMERY
F	46	1	MONTGOMERY
M	21	1	MONTGOMERY
F	19	1	PRINCE GEORGE'S
F	20	2	PRINCE GEORGE'S
F	22	3	PRINCE GEORGE'S
F	23	1	PRINCE GEORGE'S
F	24	1	PRINCE GEORGE'S
F	25	1	PRINCE GEORGE'S
F	30	1	PRINCE GEORGE'S
F	31	1	PRINCE GEORGE'S
F	32	1	PRINCE GEORGE'S
F	35	1	PRINCE GEORGE'S
M	20	1	PRINCE GEORGE'S
M	24	1	PRINCE GEORGE'S
M	26	1	PRINCE GEORGE'S
M	27	2	PRINCE GEORGE'S
M	28	1	PRINCE GEORGE'S
M	30	1	PRINCE GEORGE'S
M	35	1	PRINCE GEORGE'S
M	36	1	PRINCE GEORGE'S
M	46	1	PRINCE GEORGE'S

F	18	1	WICOMICO
F	20	1	WICOMICO
F	31	1	WICOMICO
M	21	1	WICOMICO
M	27	1	WICOMICO
M	37	1	WICOMICO
F		1	BALTIMORE CITY
F	19	1	BALTIMORE CITY
F	20	2	BALTIMORE CITY
F	22	1	BALTIMORE CITY
F	28	1	BALTIMORE CITY
F	35	1	BALTIMORE CITY
F	36	1	BALTIMORE CITY
F	40	1	BALTIMORE CITY
F	42	1	BALTIMORE CITY
F	46	1	BALTIMORE CITY
F	48	1	BALTIMORE CITY
M	17	1	BALTIMORE CITY
M	19	1	BALTIMORE CITY
M	20	2	BALTIMORE CITY
M	21	1	BALTIMORE CITY
M	22	1	BALTIMORE CITY
M	23	3	BALTIMORE CITY
M	25	3	BALTIMORE CITY
M	28	1	BALTIMORE CITY
M	31	1	BALTIMORE CITY
M	35	1	BALTIMORE CITY
M	60	1	BALTIMORE CITY
M	69	1	BALTIMORE CITY
U	22	1	BALTIMORE CITY
U	31	1	BALTIMORE CITY
U	37	1	BALTIMORE CITY
SUBTOTAL		89	
TOTAL		117	

VIRAL HEPATITIS

ORGANISM	# OF SPECIMENS	POSITIVES	JURISDICTION
HEPATITIS A			
	2	0	ALLEGANY
	2	0	ANNE ARUNDEL
	18	0	BALTIMORE
	3	0	BALTIMORE CITY
	1	0	CALVERT
	1	0	CARROLL
	1	0	CECIL
	4	0	HARFORD
	1	0	HOWARD
	2	0	MONTGOMERY
	3	0	PRINCE GEORGE'S
	1	0	SAINT MARY'S
SUBTOTAL	39	0	
HEPATITIS B			
	125	0	ALLEGANY
	92	2	ANNE ARUNDEL
	92	10	BALTIMORE
	929	10	BALTIMORE CITY
	5	0	CALVERT
	58	0	CARROLL
	151	3	CECIL
	2	0	CHARLES
	105	4	FREDERICK
	8	0	GARRETT
	94	0	HARFORD
	40	2	HOWARD
	7	0	KENT
	353	1	MONTGOMERY
	588	13	PRINCE GEORGE'S
	1	0	QUEEN ANNE'S
	3	0	SAINT MARY'S
	8	0	SOMERSET
	8	0	TALBOT
	2	0	UNKNOWN
	35	1	WASHINGTON
	165	0	WICOMICO
SUBTOTAL	2,871	36	
HEPATITIS C			
	126	10	ALLEGANY
	102	23	ANNE ARUNDEL
	87	10	BALTIMORE
	365	145	BALTIMORE CITY
	5	0	CALVERT
	59	13	CARROLL
	77	10	CECIL
	2	0	CHARLES
	113	6	FREDERICK
	9	0	GARRETT
	33	3	HARFORD
	13	1	HOWARD
	7	2	KENT
	40	4	MONTGOMERY
	290	6	PRINCE GEORGES
	1	0	QUEEN ANNE'S
	2	0	SAINT MARY'S
	3	0	SOMERSET
	6	1	TALBOT
	2	1	UNKNOWN
	12	0	WASHINGTON
	43	1	WICOMICO
SUBTOTAL	1,397	236	
TOTALS	4,307	272	

CRITICAL LINK

If you prefer to receive your issues of *Critical Link* electronically, send your address to:

criticallink@dhmh.state.md.us

It is also available on line at

<http://www.dhmh.state.md.us/labs/html/critical-link.html>

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RABIES

SOURCE	#	JURISDICTION
BAT	1	BALTIMORE
	1	WICOMICO
	1	BALTIMORE CITY
	1	BALTIMORE
CAT	1	CALVERT
	1	CECIL
	2	FREDERICK
	1	HARFORD
	1	HOWARD
	1	CHARLES
FOX	1	MONTGOMERY
	2	PRINCE GEORGE'S
	1	WORCESTER
	1	ANNE ARUNDEL
RACCOON	1	BALTIMORE
	1	CALVERT
	1	CARROLL
	1	CECIL
	2	FREDERICK
	2	HARFORD
	1	HOWARD
	4	MONTGOMERY
	4	PRINCE GEORGE'S
	1	ST. MARY'S
	2	SOMERSET
1	TALBOT	
SKUNK	1	WASHINGTON
	1	WORCESTER
	1	ALLEGANY
	1	CAROLINE
	1	CARROLL
	1	CECIL
	2	CHARLES
	2	FREDERICK
	1	ST. MARY'S
	TOTAL POSITIVES	48
TOTAL SPECIMENS	375	

CHLAMYDOPHILIA (CHLAMYDIA) PSITTACI

REPORTED QUARTERLY
NO REPORT THIS MONTH

CD4 FLOW CYTOMETRY WORKLOAD

REPORTED QUARTERLY
NO REPORT THIS MONTH

NEWBORN & CHILDHOOD SCREENING

STATISTICS FOR OCTOBER 2007

PRESUMPTIVE POSITIVES

DISORDERS	#
PHENYLKETONURIA	4
MAPLE SYRUP URINE DISEASE	2
HOMOCYSTINURIA	8
TYROSINEMIA	7
ARGININEMIA	1
CITRULLINEMIA	0
GALACTOSEMIA	0
BIOTINIDASE DEFICIENCY	0
HYPOTHYROIDISM	104
HEMOGLOBIN -DISEASE	35
HEMOGLOBIN -BENIGN	393
CONGENITAL ADRENAL HYPERPLASIA (CAH)	36
CYSTIC FIBROSIS	0
FATTY ACID OXIDATIONS	10
ORGANIC ACIDEMIAS	22
ACYLCARNITINE - BORDERLINE	5
ACYLCARNITINE - OTHERS	8

MONTHLY TOTALS

# OF SPECIMENS SCREENED	11,779
NUMBER OF TESTS	822,952
% OF UNSATISFACTORY SPECIMENS	4.59

YEAR-TO-DATE CONFIRMED CASES

CONDITIONS	# CONFIRMED
MCAD	2
3MCC	3
SCAD	2
VLCAD	2
GA-I	0
3-HYDROXY 3 METHYLGLUTARYL COA LYASE DEFICIENCY (HMG)	1
MAPLE SYRUP URINE DISEASE	2
PKU- CLINICALLY SIGNIFICANT	2
VARIATANT HYPERPHENYLALANINEMIA - NOT CLINICALLY SIGNIFICANT	2
GALACTOSEMIA- CLASSICAL GALT DEFICIENCY	3
GALACTOSEMIA - VARIANT	1
BIOTINIDASE DEFICIENCY	2
GALACTOSE EPIMERASE DEFICIENCY	0
GALACTOSE TRANSFERASE DEFICIENCY- (GALT CLASSICAL)	1
PARTIAL BIOTINIDASE DEFICIENCY	2
CAH- CLASSICAL SALT WASTING	2
CAH-NON-CLASSICAL	0
HYPOTHYROIDISM - PRIMARY	7
HYPOTHYROIDISM - SECONDARY	2
OTHER HYPOTHYROIDISM	6
TRANSIENT HYPOTHYROIDISM	1
SICKLE CELL DISEASE -SS	10
SICKLE CELL DISEASE -SC	5
SICKLE CELL DISEASE -S BETA THALASSEMIA	2
CYSTIC FIBROSIS	8

HIV ANTIBODY SCREENING – BLOOD (OCTOBER 2007)

SPECIMEN SOURCES	TOTAL	POSITIVE EIA	%	POSITIVE WB	%
HEALTH DEPARTMENTS AND CLINICS	2,571	128	4.98%	115	89.84%
HOSPITALS	143	5	3.50%	5	100.00%
DETENTION CENTERS	661	8	1.21%	5	62.50%
PRIVATE PHYSICIANS	8	0	0.00%	0	0.00%
STUDENT HEALTH CLINICS	368	2	0.54%	1	50.00%
EMPLOYEE HEALTH CLINICS	21	0	0.00%	0	0.00%
AUTOPSIES	314	9	2.87%	4	44.44%
ORGAN/TISSUE DONORS	67	0	0.00%	0	0.00%
TOTAL	4,153	152	3.66%	130	85.53%

ENVIRONMENTAL CHEMISTRY

SAMPLES	# NON-COMPLIANT	# TESTED
ASBESTOS		
AIR	0	0
BULK	5	8
AIR QUALITY		
PM _{2.5}	0	465
PM ₁₀	0	0
RADIATION		
AIR/CHARCOAL FILTERS	0	72
MILK	0	4
WIPES	0	53
RAW WATER	0	12
VEGETATION	0	0
OTHER	0	0
DRINKING WATER		
METALS		
COMMUNITY	20	95
NON-COMMUNITY	1	25
PRIVATE WELLS	61	213
PESTICIDES & PCBs		
COMMUNITY	0	77
NON-COMMUNITY	0	24
PRIVATE WELLS	0	5
VOLATILE ORGANIC COMPOUNDS		
COMMUNITY	1	378
NON-COMMUNITY	0	101
PRIVATE WELLS	0	131
RADIATION		
COMMUNITY	30	71
NON-COMMUNITY	0	0
PRIVATE WELLS	5	9
INORGANICS		
COMMUNITY	0	9
NON-COMMUNITY	3	42
PRIVATE WELLS	2	347
FOOD CHEMISTRY		
SUSPECTED TAMPERING	0	0
MICROSCOPIC FILTH	0	0
LABELING	0	0
SURVEILLANCE	0	0
CHEMICAL CONTAMINATION	0	2
TOTAL	128	2,143

LEAD ENVIRONMENTAL

TEST	#	ELEV	BRL	UNSAT
TOTAL PAINT	2	2	0	0
TOTAL SOIL	4	3	0	0
DUST				
FLOOR	400	30	342	2
SILL	599	29	511	0
WELL	330	22	227	0
OTHER	25	3	21	0
TOTAL DUST	1,354	84	1,101	2
TOTAL SAMPLES	1,360	89	1,101	2

INTERPRETATION OF RESULTS:

= Number of Samples Received
 ELEV= Elevated
 BRL= Below Reporting Limit
 UNSAT = Unsatisfactory
 PAINT Positive in excess of 0.5%
 SOIL Action level 400 - 5,000 ppm
 DUST Clearance limits: Floor/Other 40 ug/sq ft
 Window Sill 250 ug/sq ft
 Window Well 400 ug/sq ft

LEAD SCREENING - BLOOD LEAD

CLASS	RANGE ug/dl	# TESTED
MARYLAND		
I	<10	162
IIA	10-14	11
IIB	15-19	13
III	20-44	16
IV	45-69	2
V	>69	0
TOTAL		204
WASHINGTON DC		
I	<10	46
IIA	10-14	0
IIB	15-19	0
III	20-44	0
IV	45-69	0
V	>69	0
TOTAL		46

VIRAL LOAD SPECIMENS (OCTOBER 2007)

HIV-1 RNA Copies/ml	<10 ³	10 ³ – 10 ⁴	10 ⁴ – 10 ⁵	>10 ⁵	Totals
ALLEGANY COUNTY HEALTH DEPT.	8	1	4	1	14
BALTIMORE COUNTY HEALTH DEPT.	1	0	0	0	1
FREDERICK COUNTY HEALTH DEPT.	3	1	0	0	4
HOWARD COUNTY HEALTH DEPT.	0	2	0	0	2
MONTGOMERY COUNTY HEALTH DEPT.	90	10	9	5	114
PRINCE GEORGE'S COUNTY HEALTH DEPT.	70	11	14	7	102
WASHINGTON COUNTY HEALTH DEPT.	3	0	1	0	4
WICOMICO COUNTY HEALTH DEPT.	0	0	1	1	2
SUBTOTALS	175	25	29	14	243
DEPT. OF CORRECTIONS	75	19	35	16	145
TOTALS	250	44	64	30	388



MAILING LABEL

Maryland Department of Health & Mental Hygiene
 J. Mehsen Joseph Public Health Laboratory
 Critical Link c/o Georgia Corso L-15
 201 West Preston Street
 Baltimore Maryland 21201

