



TEXAS
Health and Human
Services

Texas Department of State
Health Services

RABIES PREVENTION IN TEXAS

[Texas Department of State Health Services](#)
[Zoonosis Control](#)

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This document can be accessed at:

<https://dshs.texas.gov/idcu/disease/rabies/information/prevention/>

**Texas Health and Human Services
Texas Department of State Health Services**

RABIES PREVENTION IN TEXAS

For assistance on problems or questions regarding rabies prevention, call your local health department, a Zoonosis Control regional office (contact information listed on pages 26-29 of this document) of the Texas Department of State Health Services (DSHS), the DSHS Zoonosis Control Branch (512-776-7255 or 512-776-7111), or the DSHS Infectious Disease Control Unit (512-776-7455) during working hours. For emergency consultations on nights, weekends, and holidays, contact the DSHS Public Health Region (PHR) for your area:

| PHR | PHR EMERGENCY HOTLINE |
|------------|------------------------------|
| 1 | 806-744-3577 |
| 2/3 | 817-825-9230 |
| 4/5N | 866-310-9698 |
| 6/5S | 800-270-3128 or 713-767-3000 |
| 7 | 254-778-6744 |
| 8 | 210-949-2121 |
| 9/10 | 888-847-6892 |
| 11 | 956-421-5559 |

INTRODUCTION

Although rabies rarely affects humans in the United States, every year many people receive rabies postexposure prophylaxis (PEP). Appropriate management of those who may have been exposed to rabies infection depends on the evaluation of the risk of infection (type of exposure, type of animal, availability and rabies vaccination status of the animal involved in the exposure, etc.) and the efficacy and risk of prophylactic treatment. All available methods of systemic prophylactic treatment possibly can be complicated by instances of adverse reactions. These are rarely severe. Decisions on management must be made promptly; the longer treatment is postponed, the less likely it is to be effective. The urgency for treatment must be tempered by recognition that human rabies is an extremely rare event in Texas and that hasty decisions have led to the inappropriate vaccination of people who were not at risk for infection.

Data on the efficacy of active and passive immunization after rabies exposure have come from both human and animal studies. Evidence from laboratory and field experience in many areas of the world indicates that PEP (combining wound cleansing, vaccine, and rabies immune globulin) is uniformly effective when appropriately used. Rabies has not occurred in persons who have received prompt PEP following the guidelines found in this manual.

Since the mid-1900s, the number of human rabies cases has declined significantly in the United States. This is probably due to several factors. Improved domestic animal control (including effective leash laws, domestic animal rabies vaccination programs, and stray animal collection) has been a major factor. Before the implementation of improved animal control measures, rabid domestic animals were the most common source of human rabies in the United States. They remain the leading source of human rabies in many other parts of the world.

The control of two recent major rabies epizootics in Texas has also contributed to the low incidence of rabies in domestic animals. The aggressive use of oral rabies vaccination programs for coyotes and gray foxes, which began in selected strategic areas of Texas during 1995, has greatly decreased the risk of domestic animals and humans becoming infected with the rabies virus via these wildlife species.

Rabies continues to be enzootic in skunks in Texas. The number and percentage of skunks that are positive for rabies varies cyclically. Field studies have been conducted on oral rabies vaccination efforts for skunks.

Rabies is also enzootic in bats in Texas. In the United States, the vast majority of recent human rabies cases have been identified as being due to a bat variant of rabies virus, often with the victim not remembering or disclosing an animal bite of any kind. This has resulted in a more liberal interpretation of rabies exposure by this mammal:

“...in situations in which a bat is physically present and the person(s) cannot reasonably exclude the possibility of a bite exposure, post exposure prophylaxis should be given unless prompt capture and testing of the bat has excluded rabies virus infection.” (MMWR 1996; 45:209)

RABIES BIOLOGICALS

There are two types of rabies immunizing products for use in humans: 1) vaccines that induce an active immune response, which requires about 7-10 days to develop but may persist for as long as a year or more; and 2) immune globulins that provide rapid passive immune protection for a short period of time (half-life of approximately 21 days). Both types of products should be used concurrently for PEP in those persons who have never received prior vaccination against rabies.

Vaccines for Use in Texas

This section contains some pertinent information on some of the rabies biologicals that may be available in Texas. The package insert should be consulted before the use of any of these products.

HDCV

Human diploid-cell vaccine (HDCV): HDCV is prepared using a rabies virus grown in human diploid cell culture, which is then inactivated. Vaccine is supplied as 1 ml, single-dose vials of freeze-dried vaccine with accompanying diluent for intramuscular (IM) injection for pre- and postexposure administration. It must be used immediately after reconstitution.

PCECV

Purified chick embryo cell vaccine (PCECV): PCECV contains a freeze-dried inactivated rabies virus grown on cultures of chicken fibroblasts. This vaccine is licensed in the United States for IM use in both pre- and postexposure immunization. The schedules and dosage for PCECV vaccine are the same as for HDCV.

HRIG

Human rabies immune globulin (HRIG): HRIG is concentrated rabies antibodies collected from the plasma of immunized human donors. Two different concentrations of HRIG are available in products approved for the United States: 150 IU/ml and 300 IU/ml. Calculate the dosage carefully based on the patient's current body weight and the concentration of the specific product used.

RATIONALE OF TREATMENT

The physician must evaluate each possible rabies exposure. Local or state public health officials should be consulted if questions arise about the need for prophylaxis. Additionally, a helpful guide entitled *Human Rabies Postexposure Prophylaxis (PEP) in Texas – Guide (Table 1)* can be found in the **RABIES POSTEXPOSURE PROPHYLAXIS (PEP) FOR HUMANS** section of this manual.

MANAGEMENT OF BITING ANIMALS

Per state law, all animal bites to a human must be reported to the local rabies control authority (LRCA), which may be animal control, the sheriff, or another local authority. A healthy dog, cat, or domestic ferret that has potentially exposed a person to rabies should be quarantined in a licensed facility or a veterinary clinic (under specific conditions defined in law, home confinement may be allowed by the LRCA), observed for 10 days from the time of the exposure, and evaluated by a veterinarian at the first sign of illness during the observation period. Any illness in the animal should be reported immediately to the LRCA. If signs suggestive of rabies develop, the animal should be euthanized in a manner that preserves the head in an undamaged state and its head removed and shipped at 32 to 45 degrees Fahrenheit for examination by a qualified laboratory designated by the DSHS. Any unowned dog, cat, or domestic ferret that potentially exposes a person to rabies may be humanely killed and the head submitted for rabies examination.

Signs of rabies in wild animals cannot be interpreted reliably; therefore, any free-roaming high-risk animal* that potentially exposes a person to rabies must be euthanized at once (without damage to the head) and the brain submitted for examination for rabies. A low-risk animal that the LRCA has cause to believe may be rabid must likewise be tested. (Refer to Table 1. for a list of high- and low-risk animals.) If the brain is negative by fluorescent-antibody examination for rabies, the saliva can be assumed to contain no virus and the exposed person need not be treated. Exotic animals, such as lions, tigers, or monkeys, that have been kept in captivity for extended periods of time are unlikely to be infected with rabies; they can be confined and observed for thirty days rather than euthanized and tested for rabies. Livestock can also be confined and observed for 30 days in lieu of testing.

* Refer to Texas Administrative Code, Sections 169.27(e) and (h) or consult with the LRCA for your area pertaining to exemptions to mandatory euthanasia for certain high-risk animals that meet captivity parameters as specified in state law:

[http://texreg.sos.state.tx.us/public/readtac\\$ext.ViewTAC?tac_view=5&ti=25&pt=1&ch=169&sch=A&rl=Y](http://texreg.sos.state.tx.us/public/readtac$ext.ViewTAC?tac_view=5&ti=25&pt=1&ch=169&sch=A&rl=Y)

COLLECTION AND SUBMISSION OF ANIMAL SPECIMENS FOR RABIES TESTING AT THE DSHS LABORATORY IN AUSTIN

When packing specimens for rabies diagnosis, some basic information, as mandated in Texas Administrative Code, Section 169.33, is provided below. For details on packing, labeling, shipping, and Code of Federal Regulations requirements, plus laboratory hours and standard procedures, please refer to the website of the DSHS Laboratory Services Section at http://www.dshs.state.tx.us/lab/rab_prep-ship.shtm

- Damage to the brain by shooting or other trauma shall be avoided.
- The head of the suspect animal shall be separated from the body by a qualified person wearing personal protective equipment as soon as possible after the death of the animal. Only the head shall be submitted with the exception that whole bats and small rodents may be submitted. If only the brain is submitted rather than the entire head, the minimum tissue requirements for rabies testing are a complete transverse cross section of the brain stem and tissue from one of the following: cerebellum and/or hippocampus. Submissions that do not meet these tissue

requirements will be considered unsatisfactory due to a lack of sufficient material. Live specimens will not be accepted. Due to biohazard safety guidelines, the laboratory is unable to return carcasses to submitters. Note: the rabies laboratory cannot accept live animals (an issue with bats in the past). The rabies laboratory has no means in place to euthanize animals. Therefore, with whole-animal specimens (bat or small rodent), submitters need to confirm that the animal is deceased before submitting it. Placing bats in the refrigerator is not a viable option for euthanasia because they go into a state of torpor to save energy when it's cold and don't necessarily die.

- The specimen shall be immediately chilled to between 32 degrees Fahrenheit and 45 degrees Fahrenheit either in a refrigerator or by packing for shipping with sufficient amounts of refrigerants in the container; the specimen should not be frozen. When shipping, sufficient refrigerant shall be added so the specimen will remain chilled for a minimum of 48 hours. Do not use dry ice. Gel packs or similar refrigerants are recommended. Ice is not recommended.

- If specimens are shipped, containment in compliance with requirements in the Code of Federal Regulations (CFR), Title 49, shall be used for packing. Packing methods shall prevent leakage and provide for proper identification (such as an identification number) of the specimen.

- A completed DSHS Form G-9, Rabies Submission Form, which is available at the department's Laboratory Services Section, DSHS, 1100 West 49th Street, Austin, Texas 78756, or at http://www.dshs.state.tx.us/lab/rab_testing.shtm, is required for each specimen submitted to the DSHS Laboratory Services Section. Each form must contain the same identification information provided with the specimen. Submission form(s) shall be contained in a water-proof bag.

- Labeling on the outside of the shipping container shall be legible and include:
 1. name, address, and telephone number of the laboratory;
 2. name, return address, and telephone number of the shipper;
 3. language in compliance with requirements in the CFR, Title 49, pertaining to the shipment of infectious substances for diagnostic purposes; and
 4. the following information: "RABIES IDENTIFICATION TEAM, LABORATORY SERVICES SECTION - REFRIGERATE ON ARRIVAL."

- The following procedures are required for shipment:
 1. shipment shall be by bus or other reliable carrier; the department does not recommend the United States Postal Service. If an overnight carrier (other than bus) is used, ship the specimen such that it will arrive by Friday or delay shipment until Monday. Do not ship via overnight carrier on Friday or the day before a holiday. These services do not deliver to the department on the weekend or on holidays;
 2. a shipping receipt will be obtained and retained by the shipper;
 3. at the time of the shipment, the shipper shall notify laboratory personnel of the shipment via telephone or laboratory-approved electronic format; and
 4. the shipper shall provide the return postage (in the form of stamps, not money) if return of the shipping container is desired.

LIST OF DSHS-DESIGNATED LABORATORIES

Contact information is provided to allow submitters to check with a specific laboratory for information on submission and testing procedures, plus any possible charges for testing, prior to submitting the specimen.

Austin - Laboratory Services Section, Department of State Health Services, 1100 West 49th Street, Austin, Texas 78756. Telephone the rabies hotline at 1-800-252-8163 for shipment notification. For all other inquiries call the rabies laboratory at: (512) 776-7595.

Email: Nachea.Qualls@dshs.texas.gov

Website: http://www.dshs.state.tx.us/lab/rab_prep-ship.shtm

Hours: 8am to 5pm Monday-Friday; no specimens received on federal holidays

If a specimen needs to be shipped to the laboratory over the weekend, the laboratory strongly encourages shipment of the specimen by bus rather than an overnight service.

Houston Health Department Bureau of Laboratory Services, 2250 Holcombe Blvd, Houston, Texas 77030.

Telephone: 832-393-3917

Email: Cynthia.Turner@houstontx.gov

Website: <http://www.houstontx.gov/health/Lab/rabies.html>

Hours: 8am-5pm, Monday-Friday; closed on City of Houston observed holidays

San Antonio Metro Health Laboratory, 2303 SE Military Dr, Bldg 533

Rm 1110, San Antonio, TX 78223

Telephone: (210) 207-8820 or (210) 207-8787, or 210-207-8747

Email: ralph.pruett@sanantonio.gov, mark.wade@sanantonio.gov or mireya.huizar@sanantonio.gov, claudia.garcia@sanantonio.gov

Website: <http://www.sanantonio.gov/Health/HealthServices/LabServices>

Hours: 7:45am- 4:30pm Monday-Friday

City of El Paso Department of Public Health Laboratory

9566 Railroad Drive

El Paso, TX 79924

Telephone: (915) 212-0438/Fax: (915) 212-0439

Email: GaribayAX@elpasotexas.gov

GonzalezIG@elpasotexas.gov

Website: <http://www.elpasotexas.gov/public-health>

Hours: 8am-5pm, Monday-Friday; closed on City of El Paso observed holidays

MANAGEMENT OF DOMESTIC ANIMALS EXPOSED TO RABID ANIMALS

ANY DOMESTIC ANIMAL THAT IS BITTEN BY, DIRECTLY EXPOSED BY PHYSICAL CONTACT WITH, OR DIRECTLY EXPOSED TO FRESH TISSUES OF A RABID ANIMAL IS REGARDED AS HAVING BEEN EXPOSED TO RABIES.

(The following paragraphs paraphrase portions of the Texas Administrative Code, Sections 169.21 – 169.34, Rabies Control and Eradication).

An animal should be considered not currently vaccinated if documentation of vaccination within the appropriate timeframe is not available or if the initial immunization was given less than 30 days previously.

Not currently vaccinated domestic animals considered to have been exposed to rabies must be euthanized **or** vaccinated against rabies immediately, placed in confinement for 90 days, and given booster vaccinations during the third and eighth weeks of isolation. For young animals, additional vaccinations may be necessary to ensure that the animal receives at least two doses at or after the age prescribed by the USDA for the vaccine administered.

Currently vaccinated domestic animals considered to have been exposed to rabies must be euthanized **or** vaccinated immediately and placed in confinement for 45 days.

These periods of confinement for domestic animals possibly exposed to the rabies virus should not be confused with the 10-day observation period for a dog, cat, or domestic ferret that has potentially exposed a human to rabies as described in the **RABIES POSTEXPOSURE PROPHYLAXIS (PEP) FOR HUMANS** section. A dog, cat, or domestic ferret exposed to a rabid animal may develop rabies long after the exposure since the incubation period for rabies can be days to years; the average incubation, though, is typically described as 3 to 8 weeks or 3 weeks to 3 months depending on the reference. A prolonged confinement is necessary to exclude the possibility of subsequent development of rabies in a dog, cat, or domestic ferret exposed to a rabid animal.

The above recommendations apply only to animals for which there is a USDA-licensed vaccine. For all other animals, refer to the latest edition of the publication titled *The Compendium of Animal Rabies Prevention and Control* by the National Association of State Health Veterinarians. No licensed vaccine is currently available for wild animals or hybrids of wild and domestic animals. The administration of a rabies vaccine in a species for which no licensed vaccine is available is at the discretion of the veterinarian; however, an animal receiving a rabies vaccine under these conditions will not be considered vaccinated against rabies virus in potential rabies exposure situations.

RABIES POSTEXPOSURE PROPHYLAXIS (PEP) FOR HUMANS

The essential components of animal bite wound management are prompt, thorough cleansing of wounds and immunization, including administration, in most instances, of both rabies immune globulin and vaccine.

Cleansing of Wounds

Wound cleansing cannot be overemphasized. Immediate and thorough washing of all bite wounds and scratches with soap and water (and an iodine-based antiseptic, if available and the person is not allergic) is a critical measure for preventing rabies. In experimental animals, simple local wound cleansing has been shown to markedly reduce the likelihood of rabies. Tetanus vaccination and measures to control bacterial infection should be provided as indicated.

Decision to Provide PEP

The decision to treat or not to treat must be based on all available information about the circumstances surrounding the exposure incident. The *Human Rabies Postexposure Prophylaxis (PEP) in Texas - Guide (Table 1)* found in this section is helpful in evaluating a possible rabies exposure and determining whether PEP is needed.

For animals that are not free-roaming high-risk animals and are not exhibiting signs of rabies at the time of the exposure, it is typically acceptable to delay decisions about PEP for a reasonable risk-weighted period, determined through consult and risk assessment with a public health professional, to allow the designated local authority (for example, the LRCA, animal control, or sheriff) time to locate the animal for quarantine/confinement or testing.

A 10-day observation period from the time of the bite is necessary for a dog, cat, or domestic ferret that has bitten or otherwise potentially exposed a person to rabies; the animal is placed in quarantine (or home confinement, if

applicable and approved by the LRCA) until the end of this period. In dogs, cats, and domestic ferrets, clinical signs of rabies develop within 10 days (usually less) of starting to shed virus in the saliva. Therefore, if a dog, cat or domestic ferret is alive and clinically normal after a 10-day observation period following a bite, the animal could not have exposed the bite victim to the rabies virus at the time of that specific bite incident and PEP is not warranted.

Local or state health departments may be consulted to clarify the guide and to provide information concerning the prevalence of animal rabies in the geographic locale where the possible exposure occurred. Information on the number of cases of laboratory-confirmed rabies in Texas by county and species of animal is available at:

<https://dshs.texas.gov/IDCU/disease/rabies/cases/statistics.aspx>

PEP Regimen

The PEP should include administration of both rabies immune globulin (HRIG) and vaccine (such as HDCV or PCECV). An exception is made for exposed persons who have been previously immunized with the recommended pre- or postexposure regimens of HDCV or PCECV (or who have been immunized with other types of rabies vaccines and have documented rabies antibody production). In these cases, HRIG should not be given and a dose of vaccine should be given on day 0 and day 3 (**Table 2**).

The combination of rabies immune globulin and rabies vaccine is recommended for both bite exposures and nonbite exposures, regardless of the interval between exposure and treatment. **The sooner treatment is begun after exposure, the better the chance of effectiveness.** In most cases, it is acceptable to withhold PEP for up to 72 hours while awaiting rabies test results or making efforts to locate the biting animal for testing or quarantine/observation; however, if the animal was displaying clinical signs of rabies, the exposed individual should begin treatment without awaiting test results (treatment can be discontinued if test results are negative). If there was a delay in recognizing a rabies exposure, treatment should be started even if months have lapsed since that exposure. The incubation period of rabies in humans can be days to years, but it is typically described as 3 to 8 weeks or 3 weeks to 3 months depending on the reference. The incubation period can vary due to factors such as: the age and immune status of the host; quantity of rabies virus inoculated; type and depth of the bite wound; how long the virus replicates in the host tissues; distance of the bite site from the central nervous system (CNS); and time it takes for the virus to travel from the peripheral nerves to the CNS.

Table 1. Human Rabies Postexposure Prophylaxis (PEP) in Texas – Guide

These guidelines can help determine if PEP is needed after a potential rabies exposure. **Consultation with a public health professional to assess the rabies risk is generally recommended prior to making decisions about the need for PEP.** An exposure is defined as 1) an animal bite (or scratch) that breaks the skin or 2) exposure of broken skin (bled or had serous drainage within the past 24 hours) or mucous membranes to saliva, neural tissue, or tears. Stool, blood, urine, and skunk spray do not contain rabies virus. Note: per state law, all animal bites to humans must be reported to the local rabies control authority (LRCA), which may be animal control, the sheriff, or another designated local authority.

| Risk Category of Exposing Animal | Animal Availability | Quarantine with Observation | Laboratory Testing | Laboratory Testing Result | Human PEP Recommendation ¹ |
|---|---------------------|---|---|--|--|
| Low (Rabbits, opossums, armadillos, shrews, and rodents, such as mice, rats, squirrels, gophers, prairie dogs, nutria, and beavers ²) | Available | Not required | Testing is not required unless the LRCA or physician has cause to believe that the animal is rabid. | Positive | Administer PEP. |
| | | | | Non-negative ³ | PEP may be indicated. Consult public health professional for rabies risk assessment. |
| | | | | Negative | PEP not indicated. |
| | Not available | N/A | N/A | N/A | PEP may be indicated. Consult public health professional for rabies risk assessment. |
| High⁴ (Bats ⁵ , coyotes, foxes, raccoons, and skunks or type of biting animal is unknown) | Available | Not allowed for free-roaming animals ⁴ | Test animal (Consult with LRCA if the animal is not defined as free-roaming.) | Positive or non-negative ³ | Administer PEP. |
| | | | | Negative | PEP not indicated. |
| | Not available | N/A | N/A | N/A | Administer PEP. |
| Dog, Cat, Domestic Ferret⁶ | Available | Quarantine or home confinement ⁷ until the end of a 10-day observation is allowed if animal is not exhibiting signs of rabies. | Test animal if it is exhibiting signs of rabies at the time of the exposure or if it develops signs of rabies or dies during the 10-day observation period. | Positive | Administer PEP. |
| | | | | Non-negative ³ | PEP may be indicated. Consult public health professional for rabies risk assessment. |
| | | | | Negative (or animal successfully completes quarantine or home confinement) | PEP not indicated. |
| | Not available | N/A | N/A | N/A | PEP may be indicated. Consult public health professional for rabies risk assessment. |
| All Other Mammals | Available | Confinement until the end of a 30-day observation period ⁸ is allowed if animal is not exhibiting signs of rabies. | Test animal if it is exhibiting signs of rabies at the time of the exposure or if it develops signs of rabies or dies during the 30-day observation period. | Positive | Administer PEP. |
| | | | | Non-negative ³ | PEP may be indicated. Consult public health professional for rabies risk assessment. |
| | | | | Negative (or animal successfully completes confinement) | PEP not indicated. |
| | Not available | N/A | N/A | N/A | PEP may be indicated. Consult public health professional for rabies risk assessment. |

*Footnotes listed on the next page.

1. For animals that are not free-roaming high-risk animals and are not exhibiting clinical signs of rabies at the time of the potential exposure, it is typically acceptable to delay decisions about PEP for a reasonable risk-weighted period, determined through consult with a public health professional, to allow the designated local authority, such as the LRCA, animal control, or sheriff, time to locate the animal for quarantine/confinement or testing.
2. Larger types of rodents (for example, beavers) may pose more of a rabies risk than small rodents.
3. "Non-negative" includes all specimens not suitable for testing (destroyed, decomposed, etc.).
4. Refer to Texas Administrative Code, Sections 169.27(e) and (h) or consult with the LRCA pertaining to exemptions to mandatory euthanasia and testing for certain high-risk animals that meet captivity parameters as specified in state law, in which case a 30-day observation period may be applicable in lieu of euthanasia and testing:
[https://texreg.sos.state.tx.us/public/readtac\\$ext.TacPage?sl=R&app=9&p_dir=&p_rloc=&p_tloc=&p_ploc=&pg=1&p_tac=&ti=25&pt=1&ch=169&rl=27](https://texreg.sos.state.tx.us/public/readtac$ext.TacPage?sl=R&app=9&p_dir=&p_rloc=&p_tloc=&p_ploc=&pg=1&p_tac=&ti=25&pt=1&ch=169&rl=27)
5. In incidents involving bats, PEP may be appropriate even in the absence of demonstrable bite, scratch, or mucous membrane exposure in situations in which there is reasonable probability that such exposure may have occurred (e.g., sleeping individual awakes to find a bat in the room, a person witnesses a bat in the room with a previously unattended child, mentally challenged person, or intoxicated individual, etc.).
6. The decision of whether a dog, cat, or domestic ferret should be quarantined or euthanized and tested rests with the LRCA.
7. The LRCA may allow home confinement if parameters specified in the Texas Administrative Code, Section 169.27(a) have been met:
[https://texreg.sos.state.tx.us/public/readtac\\$ext.TacPage?sl=R&app=9&p_dir=&p_rloc=&p_tloc=&p_ploc=&pg=1&p_tac=&ti=25&pt=1&ch=169&rl=27](https://texreg.sos.state.tx.us/public/readtac$ext.TacPage?sl=R&app=9&p_dir=&p_rloc=&p_tloc=&p_ploc=&pg=1&p_tac=&ti=25&pt=1&ch=169&rl=27)
8. The LRCA may authorize a 30-day observation period in lieu of testing.

Table 2. Rabies postexposure prophylaxis (PEP) schedule, United States

From Centers for Disease Control and Prevention. *Use of a Reduced (4-Dose) Vaccine Schedule for Postexposure Prophylaxis to Prevent Human Rabies: Recommendations of the Advisory Committee on Immunization Practices (ACIP)*. *MMWR* 2010;59 (No. RR-2): 6.

<http://www.cdc.gov/mmwr/preview/mmwrhtml/rr5902a1.htm>

| Vaccination Status | Intervention | Regimen* |
|---------------------------|-------------------------------------|--|
| Not previously vaccinated | Wound cleansing | All PEP should begin with immediate thorough cleansing of all wounds with soap and water. If available, a virucidal agent (e.g., povidone-iodine solution) should be used to irrigate the wounds. |
| | Human rabies immune globulin (HRIG) | Administer 20 IU/kg body weight. If anatomically feasible, the full dose should be infiltrated around and into the wound(s), and any remaining volume should be administered at an anatomical site (intramuscular [IM]) distant from vaccine administration. Also, HRIG should not be administered in the same syringe as vaccine. Because HRIG might partially suppress active production of rabies virus antibody, no more than the recommended dose should be administered. |
| | Vaccine | Human diploid cell vaccine (HDCV) or purified chick embryo cell vaccine (PCECV) 1.0 mL, IM (deltoid area [†]), 1 each on days 0, [§] 3, 7 and 14. [¶] |
| Previously vaccinated** | Wound cleansing | All PEP should begin with immediate thorough cleansing of all wounds with soap and water. If available, a virucidal agent such as povidone-iodine solution should be used to irrigate the wounds. |
| | HRIG | HRIG should not be administered. |
| | Vaccine | HDCV or PCECV 1.0 mL, IM (deltoid area [†]), 1 each on days 0 [§] and 3. |

* These regimens are applicable for persons in all age groups, including children.

[†] The deltoid area is the only acceptable site of vaccination for adults and older children. For younger children, the outer aspect of the thigh may be used. Vaccine should never be administered in the gluteal area.

[§] Day 0 is the day dose 1 of vaccine is administered.

[¶] For persons with immunosuppression, rabies PEP should be administered using all 5 doses of vaccine on days 0, 3, 7, 14, and 28.

** Any person with a history of pre-exposure vaccination with HDCV, PCECV, or rabies vaccine adsorbed (RVA); prior PEP with HDCV, PCECV or RVA; or previous vaccination with any other type of rabies vaccine and a documented history of antibody response to the prior vaccination.

For immunocompetent patients, four 1.0 ml doses of HDCV or PCECV should be given intramuscularly (in the deltoid region) in adults or the anterolateral thigh in infants. Although it is recommended that the same product be used for all doses, there is no evidence of a decrease in effectiveness should a different product need to be used to complete a treatment regimen. The first dose should be given **as soon as possible after exposure**; the first day of administration is considered to be day “0.” Additional doses should be given on days 3, 7, and 14 after the first dose. The spacing between doses represents the minimum interval to produce an effective antibody level as quickly as possible. The intervals should not be shortened or lengthened. If weekends or holidays preclude adhering to the schedule, lengthen an interval, but do not shorten it; it is important to obtain the first three injections within the first 14 days, but without reducing the stated interval between injections. Antibody response following the recommended vaccination regimen has been uniformly satisfactory; therefore, routine postvaccination serologic testing is not recommended by, nor available from, the DSHS.

For immunocompromised patients, five 1.0 ml doses of HDCV or PCECV should be given intramuscularly (in the deltoid region) in adults or the anterolateral thigh in infants on days 0, 3, 7, 14, and 28.

Every attempt should be made to adhere to the recommended vaccination schedules. Once vaccination is initiated, delays of a few days for individual doses are unimportant, but the effect of longer lapses of weeks or more is unknown. Most interruptions in the vaccine schedule do not require reinitiation of the entire series. For most minor deviations from the schedule, vaccination can be resumed as though the patient were on schedule. For example, if a patient misses the dose scheduled for day 7 and presents for vaccination on day 10, the day 7 dose should be administered that day and the schedule resumed, maintaining the same interval between doses. In this scenario, the remaining dose would be administered on day 17 (or days 17 and 31 for immunocompromised patients receiving a 5th dose).

The selection of sites for intramuscular injections appears to be critical for vaccine efficacy. Again, in adults and larger children, HDCV or PCECV should be given in the deltoid area. In infants and small children, the anterolateral thigh may be used. In the two laboratory-confirmed human cases of rabies following PEP with HDCV and HRIG within 24 hours, HDCV was administered in the gluteal area. Presumably, subcutaneous fat in the gluteal area may interfere with the immunogenicity of the vaccine.

The HRIG is administered only once, at the beginning of PEP, to provide passive immunity until the patient responds to the vaccine by active production of antibodies. Complete prophylaxis, including HRIG in a non-immunized person, should still be administered even if months have lapsed between the possible exposure and its recognition. If HRIG was not given when rabies vaccination was begun, it can be given up to the eighth day after the first dose of vaccine was given. From the eighth day (Day 7 of the formal treatment regimen) on, HRIG is not indicated because an antibody response to the vaccine is presumed to have occurred. For example, if the Day 0 rabies vaccine dose was given on March 10th, HRIG should not be given after March 17th.

The recommended dose of HRIG is 20 IU/kg of body weight. The dosage should be calculated carefully based on the patient’s current body weight as two different formulations of HRIG are available in the US: 150 IU/ml and 300 IU/ml. The HRIG may partially suppress the active production of antibodies; therefore, no more than the recommended dose of HRIG should be given. As much as possible of the full dose of HRIG should be thoroughly infiltrated into and

around the wound(s). Any remaining dosage of HRIG should be administered IM in the closest muscle mass of suitable size to accommodate the remaining volume, with the caveat that it should not be administered in the same syringe or in the same anatomic site as the first vaccine dose (subsequent doses of vaccine in the series—i.e. days 3, 7, and 14—can be administered in the same anatomic location in which HRIG was administered). The HRIG should be injected into muscle, not adipose tissue. For this reason, it is recommended that HRIG not be injected into the gluteal area. If the gluteal area is used, particular care should be taken to assure IM injection. (Note: human immune globulin used to treat hepatitis cannot be substituted for HRIG.)

Additional guidelines on vaccine administration are in the *ACIP General Recommendations on Immunization* (<http://www.cdc.gov/mmwr/preview/mmwrhtml/rr6002a1.htm>).

For dealing with more significant deviations to the PEP regimen, contact the DSHS Zoonosis Control Branch (refer to contact information at the end of this document) or the Centers for Disease Control and Prevention (<https://www.cdc.gov/vaccines/vpd/rabies/hcp/index.html>).

In unusual instances, such as when the patient is immunodeficient or immunosuppressed, serologic testing (Rapid Fluorescent Focus Inhibition Test - RFFIT) is indicated 2 to 4 weeks after completing the rabies postexposure vaccination series.

These are laboratories that provide the rabies RFFIT:

Rabies Laboratory
Kansas State University
2005 Research Park Circle
Manhattan, KS 66502
Phone: 785-532-4483
Email: rabies@vet.k-state.edu
www.vet.ksu.edu/rabies

Atlanta Health Associates
309 Pirkle Ferry Rd. Suite D300
Cumming, GA 30040
Phone: 800-717-5612
Email: info@atlantahealth.net
www.atlantahealth.net

Maryland Department of Health
ATTENTION: ACCESSIONING RABIES/RFFIT TITER TESTING
1770 Ashland Ave.
Baltimore, MD 21205
Phone: 443-681-3773
<https://health.maryland.gov/laboratories/Pages/Rabies.aspx>

TREATMENT OUTSIDE THE UNITED STATES

If PEP is begun outside the United States with locally produced biologicals, it may be desirable to provide additional treatment, including restarting PEP with products licensed for use in the US, when the patient reaches the US. For specific advice in such cases, contact the Regional Zoonosis Control office for your area (refer to the Zoonosis Control contact section at the end of this document). You may also refer to the World Health Organization (<http://www.who.int/topics/rabies/en/>) or the Centers for Disease Control and Prevention (<https://www.cdc.gov/vaccines/vpd/rabies/index.html>).

PREEXPOSURE VACCINATION

Preexposure vaccinations are given for several reasons. First, it may provide protection to people with inapparent exposures to rabies. Second, it may protect persons whose postexposure therapy might be delayed. This is of particular importance for persons at high risk of being exposed in countries where the rabies biologicals may be difficult to obtain. Finally, although it does not eliminate the need for additional therapy after a rabies exposure, it simplifies therapy by eliminating the need for HRIG and decreasing the number of doses of rabies vaccine needed. The guidelines for evaluating the need for preexposure immunization are found in **Table 3**. The schedule for preexposure vaccinations is located in **Table 4**.

Table 3. Rabies preexposure vaccination guide

| Risk category | Nature of risk | Typical populations | Preexposure recommendations |
|--|--|---|---|
| Continuous | Virus present continuously, often in high concentrations. Aerosol, mucous membrane, bite, or nonbite exposure. Specific exposures may go unrecognized. | Rabies research lab worker,* rabies biologics production workers. | Primary course. Serologic testing every 6 months; booster vaccination if antibody titer falls below acceptable level.** |
| Frequent | Exposure usually episodic, with source recognized, but exposure may also be unrecognized. Aerosol, mucous membrane, bite, or nonbite exposure. | Rabies diagnostic lab workers,* spelunkers, veterinarians and staff, veterinary students, and animal control and wildlife workers in rabies-enzootic areas.*** | Primary course. Serologic testing every 2 years; booster vaccination if antibody titer falls below acceptable level.** |
| Infrequent (greater than population at large) | Exposure nearly always episodic with source recognized. Mucous membrane, bite, or nonbite exposure. | Veterinarians and staff, veterinary students, and animal-control and wildlife workers in areas of low-rabies occurrence. Travelers visiting foreign areas of enzootic rabies for more than 30 days. | Primary course; no serologic testing or booster vaccination. |
| Rare (population at large) | Exposures always episodic. Mucous membrane or bite with source recognized. | US population at large, including persons in rabies enzootic areas. | No vaccination necessary. |

* Judgment of relative risk and extra monitoring of vaccination status of laboratory workers is the responsibility of the laboratory supervisor.

** Minimum acceptable antibody level is complete virus neutralization at a 1:5 serum dilution by RFFIT. Booster dose should be administered if the titer falls below this level.

*** Texas is considered to be a rabies-enzootic area.

Again, preexposure vaccinations do not eliminate the need for prompt PEP following an exposure; it only reduces the extent of the postexposure regimen.

Vaccination Schedule

For preexposure immunization, three 1.0 ml injections of HDCV or PCECV should be given intramuscularly (IM) in the deltoid area, one on each day: 0, 7, and either 21 or 28 (the first day of administration is considered to be day “0”) (**Table 4**). The antibody response following the recommended vaccination regimen with HDCV or PCECV has been uniformly satisfactory; therefore, routine postvaccination serology is not necessary.

If possible, immunosuppressed patients should postpone rabies preexposure prophylaxis until the immunocompromising condition is resolved. When postponement is not possible, immunosuppressed persons who are at risk for rabies should have their virus-neutralizing antibody responses checked 2 to 4 weeks after completing the preexposure series.

Booster Doses of Vaccine

The recommended schedules for booster doses for persons at risk of rabies exposures are outlined in **Table 3**. The addresses and phone numbers of laboratories offering rabies serologic testing can be found in the **RABIES POSTEXPOSURE PROPHYLAXIS (PEP) FOR HUMANS** section.

Postexposure Therapy of Previously Immunized Persons

When an immunized person who was vaccinated according to the recommended preexposure or postexposure regimen with HDCV or PCECV, or who has previously demonstrated rabies antibody, is exposed to rabies, that person should receive two IM doses (1.0 ml each) of HDCV or PCECV, one immediately and one 3 days later (**Table 2**). The HRIG should **not** be given in these cases. If the immune status of a previously vaccinated person who did not receive the recommended HDCV or PCECV regimen is not known, full primary PEP (HRIG plus five doses of HDCV or PCECV) may be necessary.

Table 4. Rabies preexposure vaccination schedule

| Type of vaccination | Route | Regimen |
|---------------------|-------|---|
| Primary | IM | HDCV or PCECV, 1.0 ml (deltoid area), one each on days 0, 7, and 21 or 28 |
| Booster* | IM | HDCV or PCECV, 1.0 ml (deltoid area), day 0 only |

* Administration of a booster dose of vaccine depends on exposure risk category and serologic testing results as noted in Table 3.

HDCV or PCECV can be used for PEP or booster vaccinations even if another vaccine was used for the initial preexposure vaccination.

ADVERSE REACTIONS

Serious adverse reactions associated with rabies vaccines include systemic, anaphylactic, and neuroparalytic reactions. Serious adverse reactions occur at lower rates with the rabies vaccines currently used in the US than with those used previously.

HDCV

Local and systemic reactions may occur with the use of HDCV. In a study using five doses of HDCV, local reactions, such as pain, erythema, and swelling or itching at the injection site, were reported in about 25% of recipients. Mild systemic reactions, such as headache, nausea, abdominal pain, muscle aches, and dizziness, were reported in about 20% of recipients.

Up to 6% of persons receiving booster doses of HDCV may experience "immune complex-like" reactions (type III IgG mediated hypersensitivity reactions). The illness, characterized by onset 2-21 days post-vaccination, presents with generalized urticaria and may also include arthralgia, arthritis, angioedema, nausea, vomiting, fever, and malaise. No life-threatening cases of "immune complex-like" illness have been reported. This phenomenon rarely occurs with primary immunization with HDCV. The origin of these "immune complex-like" reactions has been attributed to sensitization to the beta-propiolactone-treated human serum albumin present in HDCV.

Those persons with a history of "immune complex-like" illness following HDCV and who must receive rabies vaccine boosters may be candidates for other rabies vaccines which do not have beta-propiolactone bound to human serum albumin. If this is not possible and HDCV must be given, the guidelines in the **Management of Adverse Reactions** section should be followed.

PCECV

As with HDCV, local reactions such as swelling, induration, and reddening have been associated with the administration of PCECV. Systemic allergic reactions are also possible and have been reported.

HRIG

Local pain and low-grade fever may follow receipt of HRIG. Although not reported specifically for HRIG, angioneurotic edema, nephrotic syndrome, and anaphylaxis have been reported after injection of other types of immune globulin (IG). These reactions occur so rarely that a causal relationship between IG and these reactions is not clear.

Management of Adverse Reactions

Once initiated, PEP should not be interrupted or discontinued because of local or mild systemic adverse reactions to rabies vaccine. Usually such reactions can be successfully managed with anti-inflammatory and antipyretic agents.

When a person with a history of hypersensitivity must be given rabies vaccine, antihistamines may be given. Epinephrine should be readily available to counteract anaphylactic reactions, and the person should be observed for 15 minutes after vaccination.

Serious systemic anaphylactic or neuroparalytic reactions occurring during the administration of rabies vaccine pose a serious dilemma for the attending physician. A patient's risk of developing rabies must be carefully considered before deciding to discontinue vaccination. Moreover, the use of corticosteroids to treat life-threatening neuroparalytic reactions carries the risk of inhibiting the development of active immunity to rabies. It is especially important in these cases that the serum of the patient be tested for rabies antibodies after completion of the PEP course. Advice and assistance on the management of serious adverse reactions in persons receiving rabies vaccines may be sought from the DSHS Emerging and Acute Infectious Disease Unit (512-776-7455 or 512-776-7676).

All adverse events or reactions to a rabies vaccine should be reported to the Vaccine Adverse Event Reporting System (VAERS); additional information and forms can be obtained at <http://vaers.hhs.gov/>.

PRECAUTIONS AND CONTRAINDICATIONS

Immunosuppression

Corticosteroids, other immunosuppressive agents, and immunosuppressive illnesses (such as HIV infection and cancer) can interfere with the development of active immunity and predispose the patient to developing rabies. Immunosuppressive agents should not be administered during PEP, unless essential for the treatment of other conditions. When PEP is administered to persons receiving corticosteroids or other immunosuppressive therapy, or to persons having an immunosuppressive illness, it is especially important that the person receive a 5th dose of vaccine on day 28 (if the person is not previously immunized) and that serum be tested for rabies antibody to ensure that an adequate response has developed. Information on the laboratories that offer rabies serologic testing is found in the **RABIES POSTEXPOSURE PROPHYLAXIS (PEP) FOR HUMANS** section.

Pregnancy

Fetal abnormalities have not been associated with rabies vaccination. Due to the consequences of an inadequately treated rabies exposure, pregnancy is not considered a contraindication to PEP. If a substantial unavoidable risk of exposure to rabies exists, preexposure prophylaxis may also be indicated during pregnancy.

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DISTRIBUTION POINTS FOR HUMAN RABIES BIOLOGICALS

Note: If seeking rabies biologicals, please contact the distribution points BEFORE sending anybody directly to the office listed.

PUBLIC HEALTH REGION 1

Regional Office - Lubbock
6302 Iola Avenue
Lubbock, Texas 79424
Phone: (806) 783-6422 or (806) 744-3577
Fax: (806) 783-6466
Hours: 8:00 a.m. - 5:00 p.m. Monday-Friday
Contact: Karen McDonald, MS
Regional Veterinarian – Cherissa Abdul Hamid, DVM, MPH - 806-783-6482

Regional Office - Amarillo
3407 Pony Express Way
Amarillo, Texas 79118
Phone: (806) 477-1104
Fax: (806) 373-4758
Hours: 8:00 a.m. - 5:00 p.m. Monday-Friday
Contact: Tonya Finch
Regional Veterinarian - vacant 806-783-6482

PUBLIC HEALTH REGION 2/3

Regional Office - Arlington, Public Health Region 3
Chase Bank Building, Second Floor, Ste 200
1301 South Bowen Road
Arlington, Texas 76013
Phone: (817) 264-4920
Fax: (817) 264-4925
Hours: 8:00 am - 5:00 pm Monday-Friday
Contact: Shannon Medrano
After Hours: (817) 825-9230

Regional Office – Abilene, Public Health Region 2
4601 South 1st Street, Suite L
Abilene, Texas 79605
Phone: (325) 795-5857
Fax: (325) 795-5891
Hours: 8:00 a.m. - 5:00 p.m. Monday-Friday
Contact: Nicholas Ferguson
After Hours: (325) 261-2684

Collin County Healthcare Services
825 N. McDonald, Ste. 130
McKinney, Texas 75069
Phone: (972) 548-4707
Fax: (972) 548-4436
Hours: 8:00 a.m. – 11:00 a.m. and 1:00 p.m. – 4:00 p.m. Monday - Friday
Contact: Susana Ramos
After Hours: (972) 547-5350

Denton County Health Department
535 S. Loop 288
Denton, Texas 76205
Phone: (940) 349-2909
Fax: (940) 349-2905
Hours: 8:00 a.m. – 4:30 p.m. Monday – Friday (closed 12:00 – 1:00 p.m.)
Contact: Lilly Metzler

Abilene Taylor Public Health District
850 N Sixth St
Abilene, Tx 79601
(325) 692-5600 Fax (325) 734-5370
Hours: 8:00-5:00 Monday – Friday
Contact: Brandie Walsh (325) 692-5600, ext. 4610; Sylvia Molina (325) 692-5600, ext. 4668

Brownwood-Brown County Health Department
510 East Lee
Brownwood, Texas 76801
Phone: (325) 646-0554
Fax: (325) 643-3591
Hours: 8:00 a.m. - 4:30 p.m. Monday – Friday (closed 12:00 – 1:00 p.m.)
Contact: Kori Leach, LVN

Wichita Falls – Wichita County Public Health District
1700 Third Street
Wichita Falls, Texas 76301
Phone: (940) 761-7874
Fax: (940) 761-7885
Hours: 8:00 a.m. - 5:00 p.m. Monday-Friday
Contact: Lisa Harris, RN, Clinical Supervisor (940) 761-7844

Grayson County Health Department
515 N. Walnut Street
Sherman, Texas 75090
Phone: (903) 893-0131
Fax: (903) 870-2023
Hours: 8:00 a.m. – 4:30 p.m. Monday-Friday (closed 12:00 p.m. – 1:00 p.m.)
Contact: Reta Cooksey 903-893-0131 X 1233

PUBLIC HEALTH REGION 4/5N

Regional Office
2521 West Front Street
Tyler, Texas 75702
Phone: (903) 533-5212
Fax: (903) 533-9502
Hours: 8:00 a.m. - 5:00 p.m. Monday-Friday (Consultations only. No Human Rabies Biologicals stocked or distributed.)
Contact: Alexandro Chavira (903) 533-5212

PUBLIC HEALTH REGION 6/5S

Regional Office
5425 Polk Ave, Suite J
Houston, Texas 77023
Phone: (713) 767-3300
Fax: (713) 767-3193
Hours: 8:00 a.m. - 5:00 p.m. Monday-Friday
Contact: Dawn Blackburn, BVMS, MSc, DACVPM (713) 767-3302

Beaumont City Health Department
950 Washington Blvd
Beaumont, Texas 77704
Phone: (409) 654-3625
Hours: 8:00 a.m. - 5:00 p.m. Monday-Friday
Contact: Mary Alexander, RN or Sherry Ulmer, RN

PUBLIC HEALTH REGION 7

Regional Office
2408 S. 37th Street
Temple, Texas 76504-7168
Phone: (254) 771-6784
Fax: (254) 771-2662
Hours: 8:00 a.m. - 5:00 p.m. Monday-Friday
Contact: Jay Leivdal

PUBLIC HEALTH REGION 8

Regional Office
7430 Louis Pasteur
San Antonio, TX 78229
(210) 949-2046
Fax: (210) 692-1457
Contact: Amanda Kieffer, DVM, MPH, DACVPM
E-MAIL: Amanda.Kieffer@dshs.texas.gov

San Antonio Metro Health District
210 N Mel Waiter
San Antonio, TX 78207
Phone: (210) 207-2095
Hours: 8-11:30 1:00-4:00
Contact: Lorraine Castro

Victoria County Public Health Department
2805 North Navarro
Victoria, TX 77901
Phone (361) 578-6281
Contact: DiAnna Harris, RN

Field Office – Maverick County
1593 Veterans Blvd.
Eagle Pass, Texas 78852
Phone: (830) 758-4253
Contact: Edilia Gonzales

Field Office - Val Verde County
173 Wildcat Drive
Del Rio, Texas 78840
Phone: (830) 768-2800
Contact: Jose Guerrero

Field Office – Uvalde County
112 Joe Carper Drive
Uvalde, Texas 78801
Phone: (210) 949-2048
Contact: Amanda Kieffer

PUBLIC HEALTH REGION 9/10

Ector County Health Department
Nursing Department
221 North Texas
Odessa, Texas 79761
Phone: (432) 498-4141
Hours: 8:00 a.m. - 5:00 p.m. Monday-Friday
Contact: Linda Cunha, RN

Texas Department of State Health Services Nursing Department - Alpine
205 North Cockrell Street
Alpine, Texas 79830
Phone: (432) 837-3877
Hours: 8:00 a.m. - 6:00 p.m. Monday - Friday
Contact: Vacant

Texas Department of State Health Services Nursing Department - Brady
1004 S. Bridge St.
Brady, TX. 76825
Phone: (325) 597-0550
Hours: 8:00 a.m. – 6:00 p.m. Monday-Friday
Contact: Toni Keltz, RN

PUBLIC HEALTH REGION 11

Regional Office
601 West Sesame Drive
Harlingen, Texas 78550
Phone: (956) 444-3212
Fax: (956) 444-3216
Hours: 8:00 a.m. - 5:00 p.m. Monday-Friday
Contact: Letty Tamayo – ext. 3212
After Hours: (956) 453-1049

Corpus Christi Sub-Office
5155 Flynn Parkway
Corpus Christi, Texas 78411
Fax: (361) 883-4414
Hours: 8:00 a.m. - 5:00 p.m. Monday-Friday
Contact: Rebecca Garcia (361) 878-3469; Lamar Torres (361) 878-3457; Lydia Vera (361) 878-3415;
or Laura Zepeda (361) 878-3416

Hidalgo County Health Department
1304 South 25th St.
Edinburg, Texas 78542
Phone: (956) 383-6221
Fax: (956) 318-2422
Hours: 8:00 a.m. - 5:00 p.m. Monday-Friday
Contact: Cynthia Orozco – ext 7233, Nelda Mendez – ext 7232

City of Laredo Health Department
2600 Cedar Avenue
Laredo, Texas 78040-4040
Phone: (956) 795-4906
Fax: (956) 795-4956
Hours: 8:00 a.m. - 5:00 p.m. Monday-Friday
Contact: Maria Paredes – (956) 727-6966
Pharmacy: Lucinda Delgado – (956) 795-4909 Fax: (956) 712-6000

ZOONOSIS CONTROL -
TEXAS DEPARTMENT OF STATE HEALTH SERVICES

PUBLIC HEALTH REGION 1

6302 Iola Avenue
Lubbock, Texas 79424
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PUBLIC HEALTH REGION 7

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Kelly Broussard, MPH – Epidemiologist – 6920

Danielle Bucklin, MS – Program Specialist -2279

Emily Cavazos – Public Health & Prevention Specialist -2790

Alexandra Colemere, MPH – Program Specialist - 3339

Colleen Cook, MS – Program Specialist -3520

Michelle Ellison – Staff Services Officer -6001

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Patrick Hunt - Program Specialist - 6270

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Bonny Mayes, MA – Epidemiologist – 2888

Briana O’Sullivan, MPH – Epidemiologist - 2890

Kamesha Owens, MPH – Program Specialist - 2914

Kathy Parker, MPH – Program Specialist – 2886

George Peck, MS, PhD – Entomologist - 2884

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